Time Series Transcriptomics

On Frog Embryos infected with *P. Aeruginosa*

Extracting Key Genes

and Gene Groups

SAHIL LOOMBA

Time Series Data

- Transcriptomics data for frog embryos infected with *P. aeruginosa*
  - Different infection levels
  - Different stages of development
  - Replicates

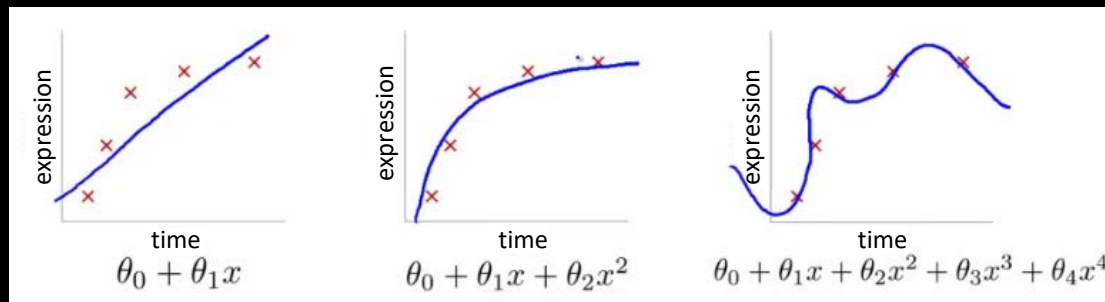| Infection (cfu)	# Replicates	Time points (day)
0	2	1, 2, 3
100	3	1, 2, 3
1000	3	1, 2, 3
10000	3	1, 2

- Probes mapped to 8726 frog genes from Xenbase

Objective

- To develop a transcriptomics model that captures key differences across treatment conditions
- Sensitive to (is a function of) time
  - Usual differential expression analysis takes place at a particular time point
- Challenges
  - Few samples per condition (6-9)
  - Even fewer samples per condition per time point (0-3)
  - Sample variance and error of measurement
- Can we still capture temporal and condition resolution?
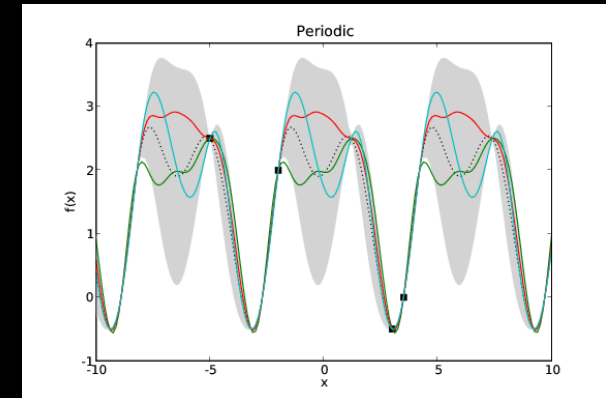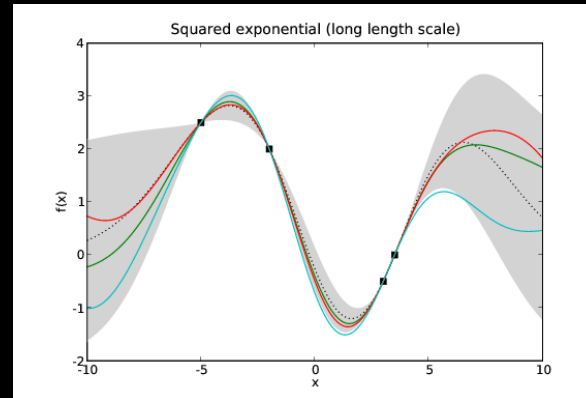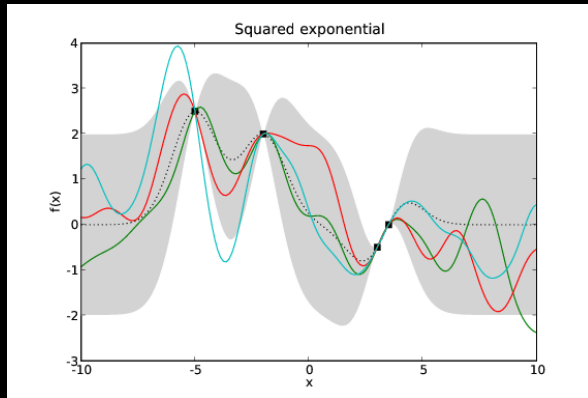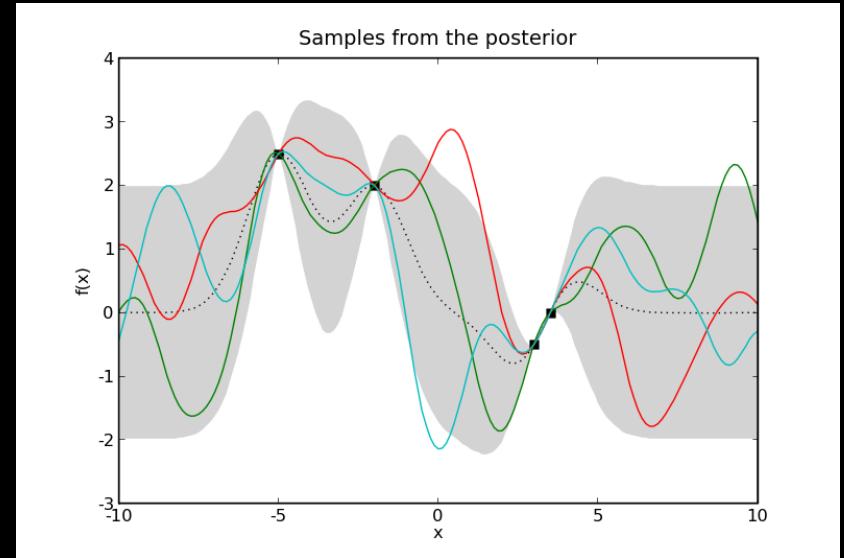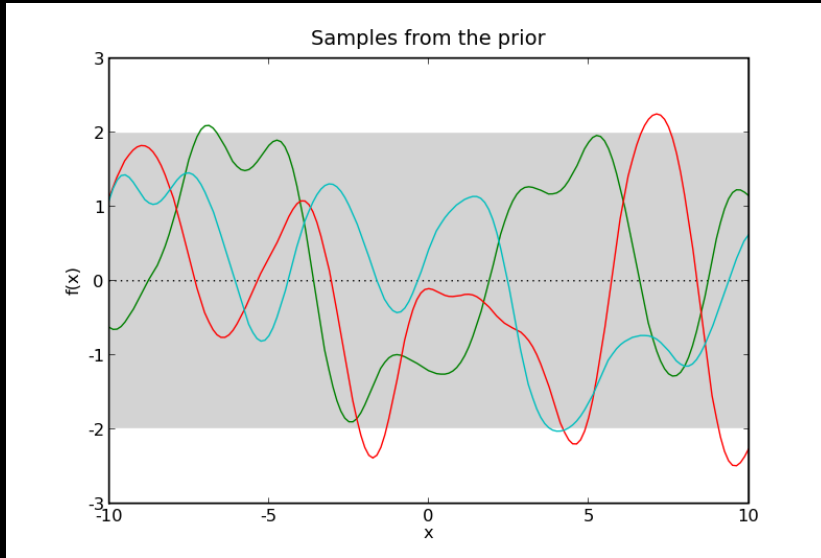 - While principally handling these issues

Gaussian Process Regression

- Regression: Given a dataset, finding a mapping from a domain to a range; usually "parametric"

- GPR: Considers the mapping between two feature spaces as a random variable itself, following a Gaussian Process prior whose covariance is a function of the observations ("non-parametric")

- If the domain space is time, it can model a time-series

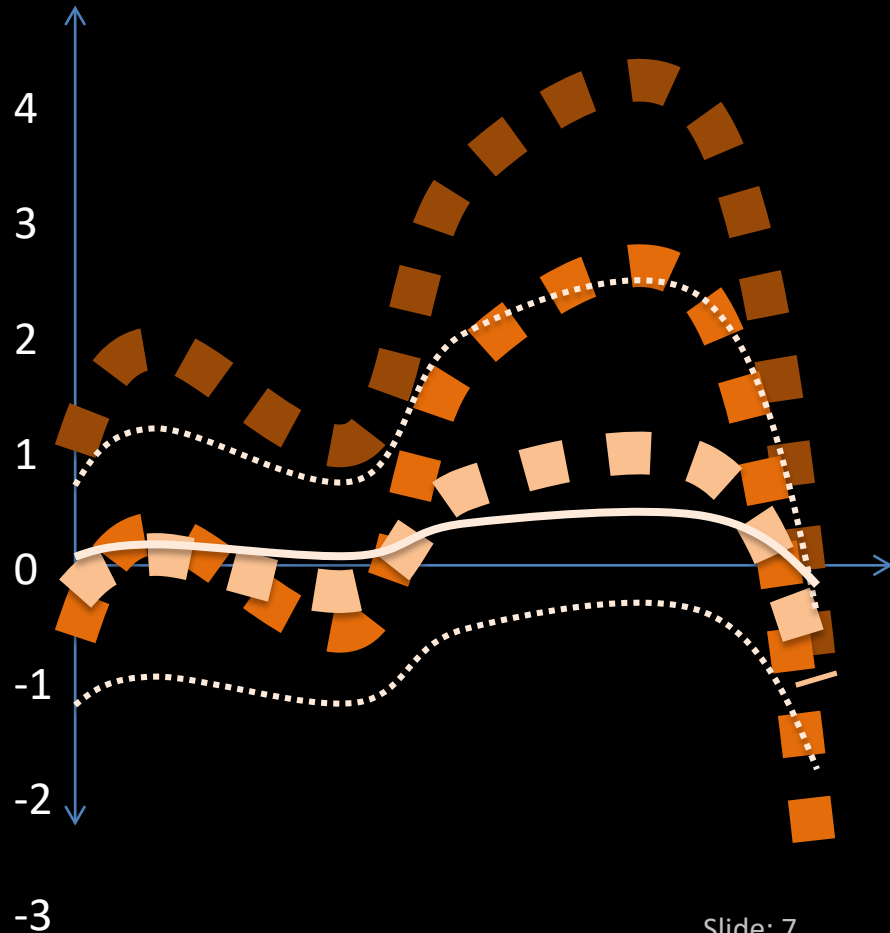# Gaussian Process Regression

Gaussian Process Regression

- No need to specify a model of regression
  - Assumption of "smoothness"
- Automatic Occam's Razor
- Makes predictions in previously unseen spaces with bounds on uncertainty of the prediction
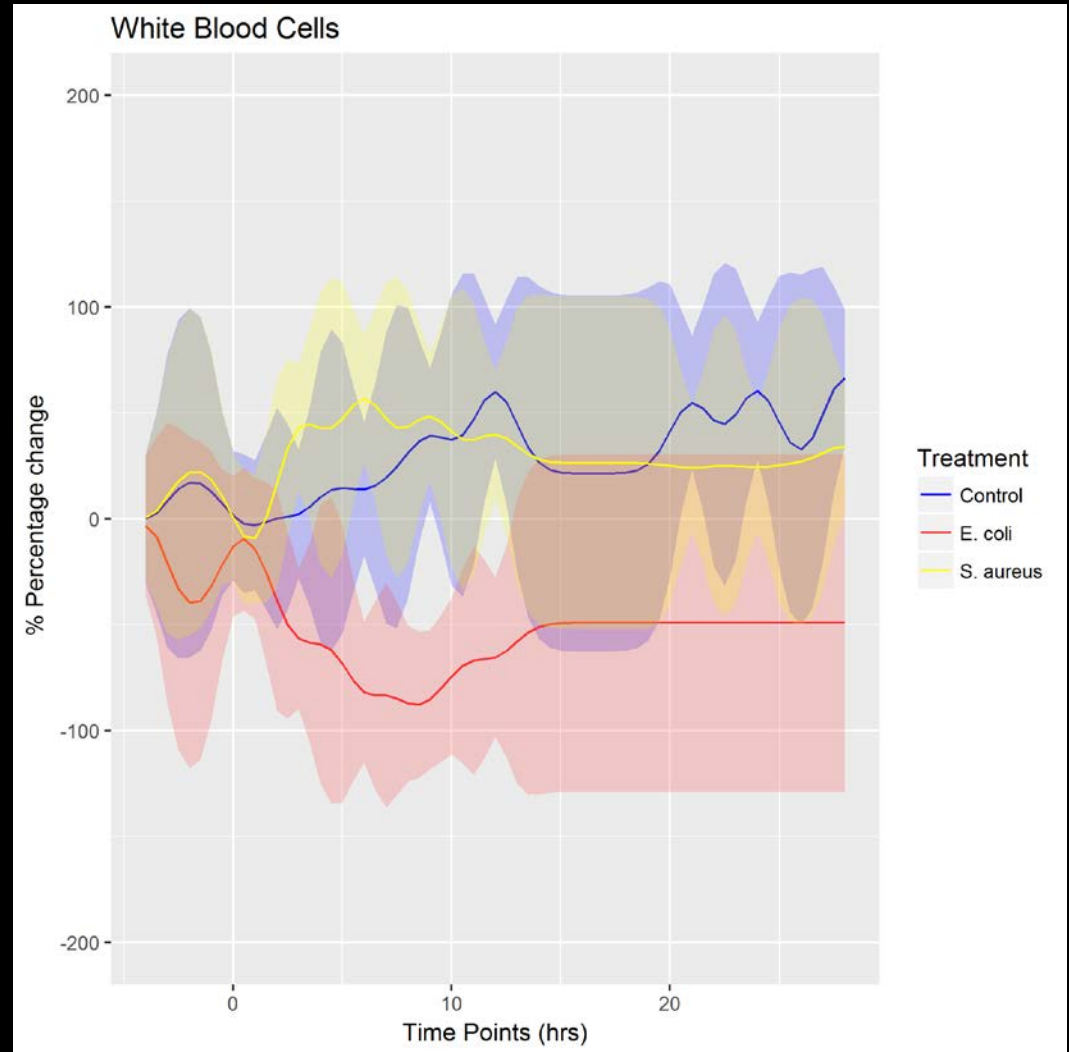
Gaussian Process Regression

- Decouples various aspects of the data:
  - Bias: *b*
  - Scale: *s*
  - Mean function: *m(x)*
  - Variance around mean function: *k(x,x')*
  - Error term: *e*

Gaussian Process Regression

For the pig studies:

Biomarkers/Physiological data for

- 15 pigs across
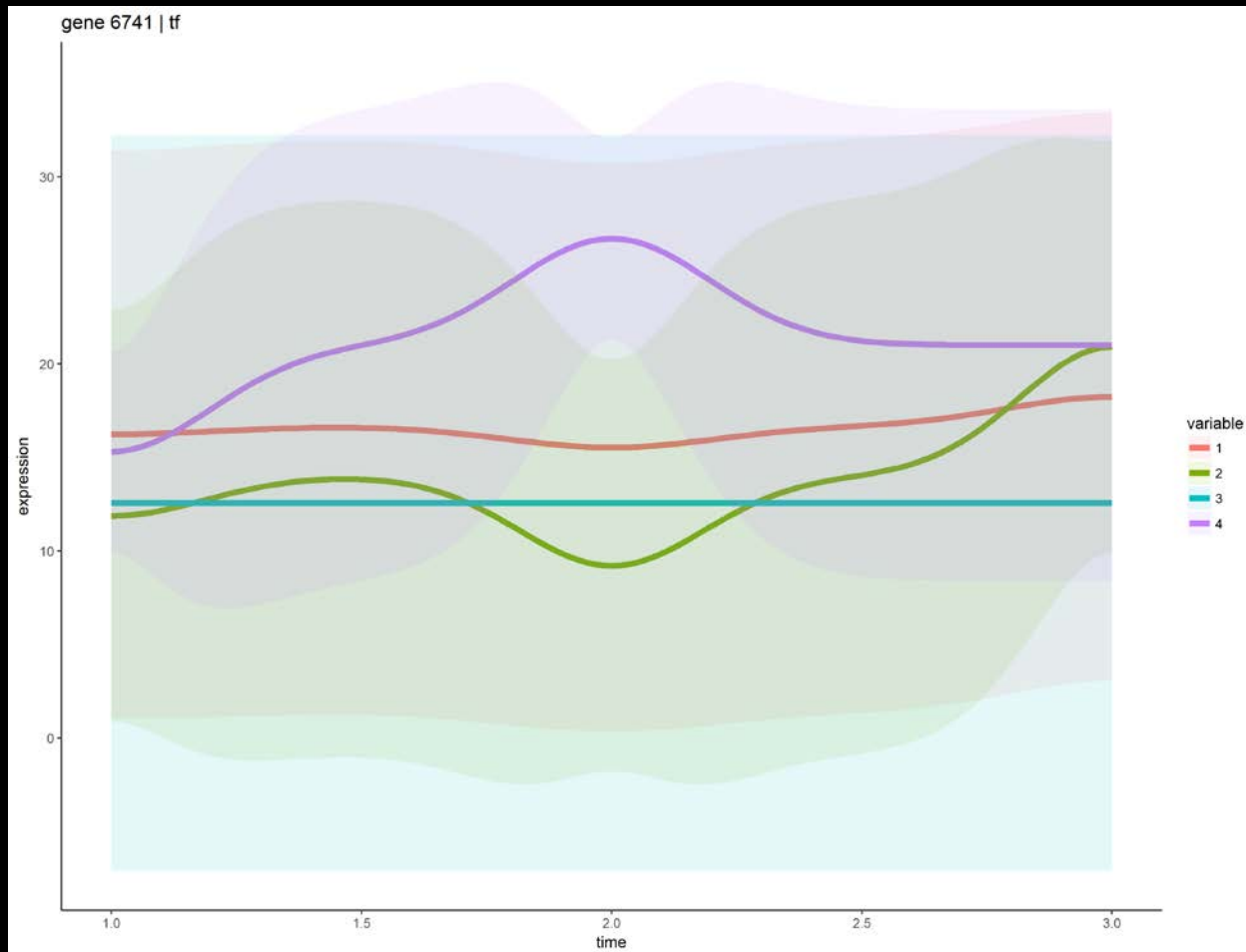- 3 conditions

White Blood Cells

GPR for Omics, Method 1

- Treat every gene (for a given condition) as an independent GP mapping from time space to expression space
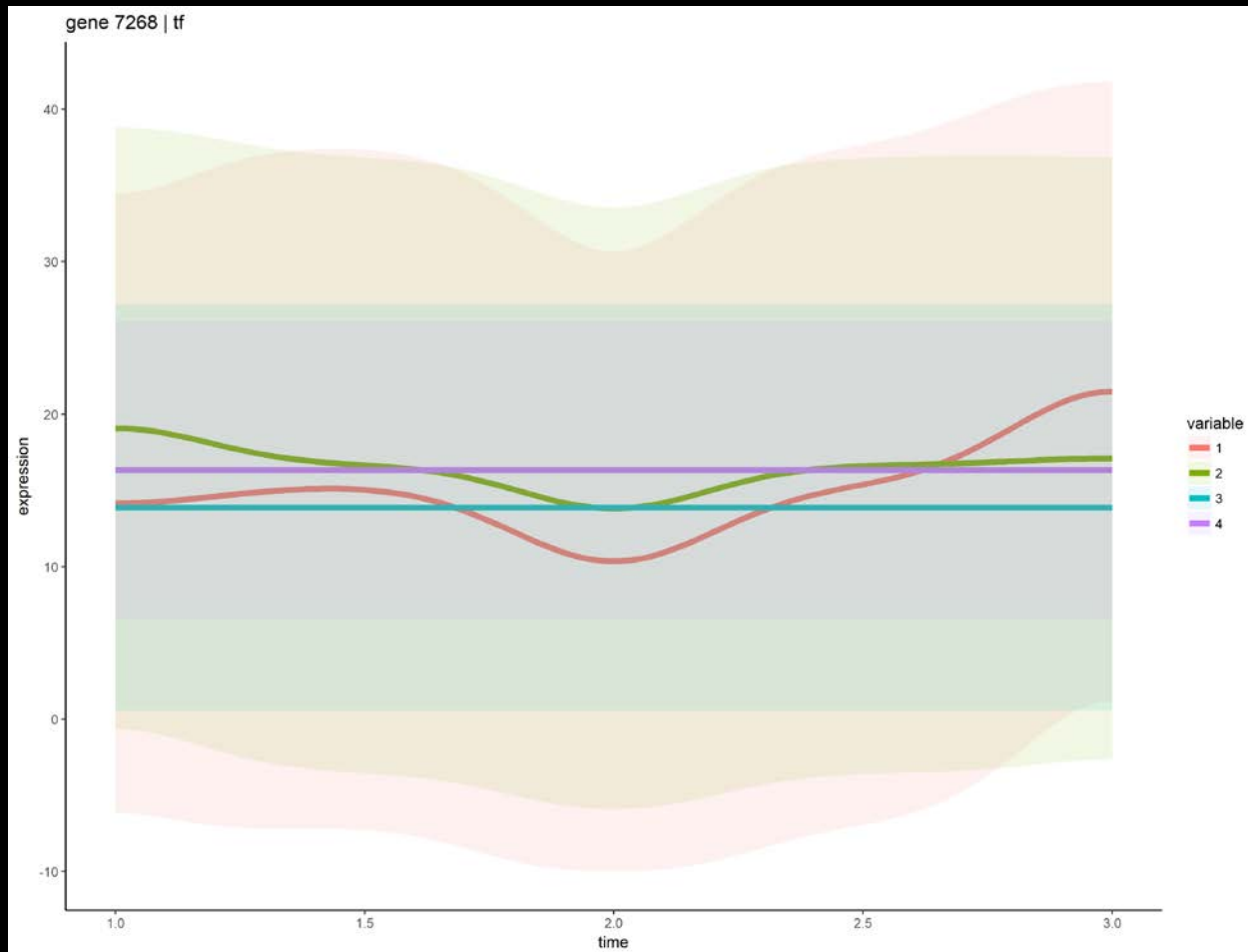
$$f : t \rightarrow g$$
$$f(t') \sim GP\big(m(t), k(t, t')\big) + \beta$$

- For every gene. learn 4 GP models corresponding to the 4 conditions

- We can plot the output of a learnt GP model by plotting its posterior mean estimate and posterior covariance estimate
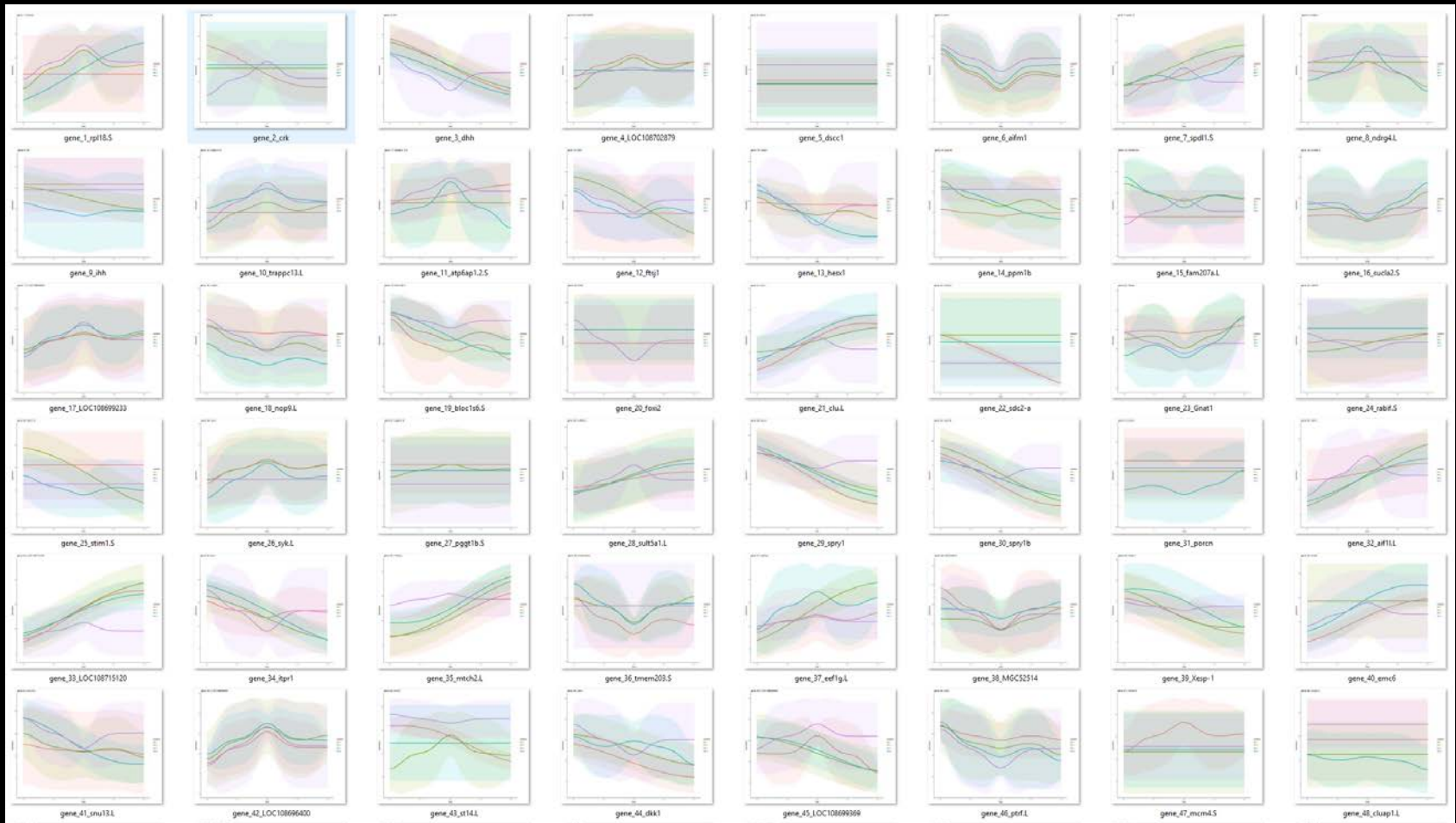
Example: Transferrin (probe1)

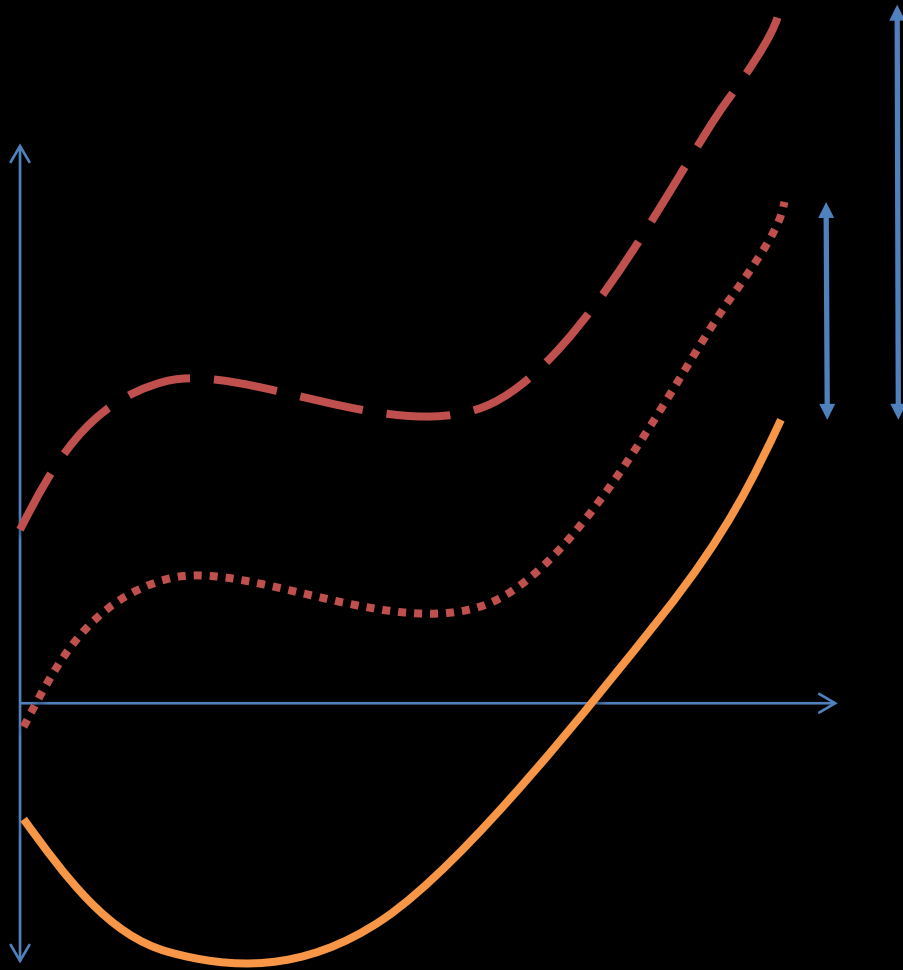# Example: Transferrin (probe2)

For all 8726 genes…
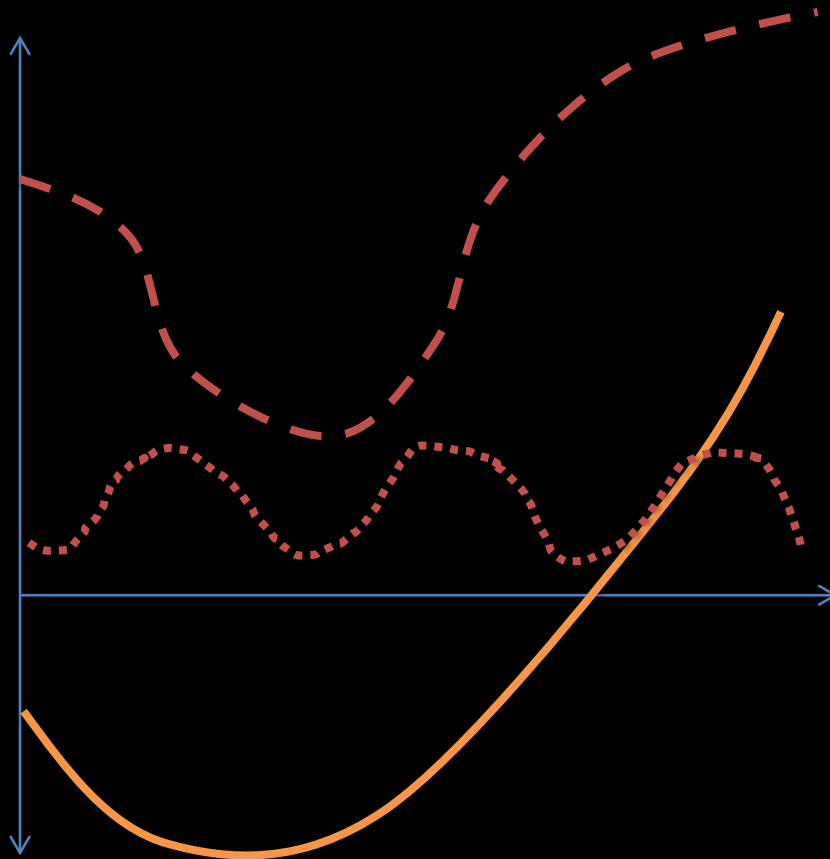
To extract key genes and gene groups...

- Visually!

- Question: How do we compare two GP models?
  - Can't compare in model space because GP is non-parametric and we have different samples to every GP

- Solution: use ideas from information theory to recharacterize every model
  - Treat output (posterior mean estimate) of learnt GP as the new "smoothed" time-series
  - Define metrics to compare those smooth series

Metric 1: (Euclidean) Distance

The further away two conditions are, the better a gene is in discriminating between them
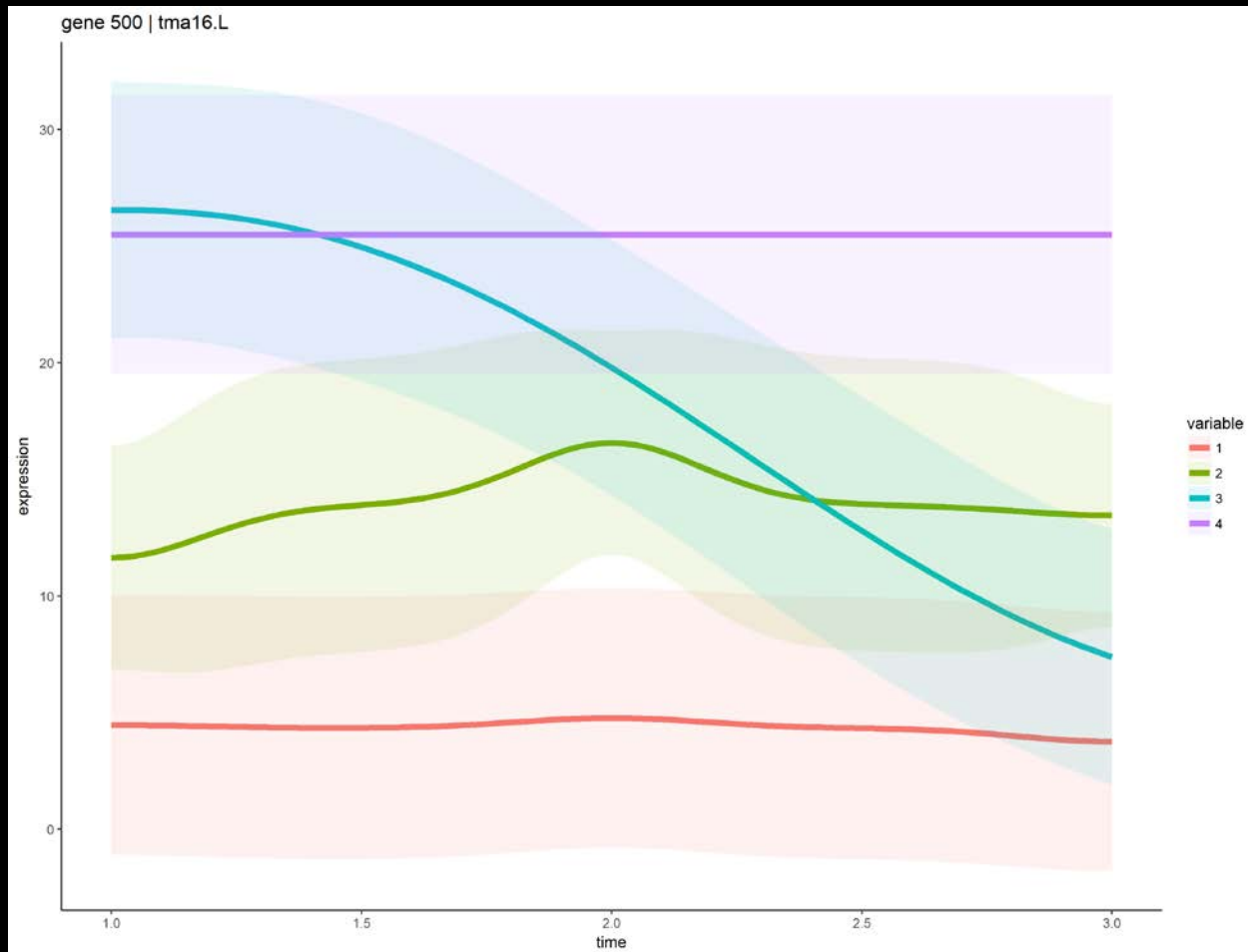
Metric 2: (Mutual) Information

The more informative (~correlated*) one condition is about the other, the more it matters

*Unlike correlation, mutual information doesn't assume linearity; more so a notion of predictability of one time-series from the other

Definition of Key Genes

- Which induce maximum sum of pairwise distances between all informative conditions

- We rank genes in decreasing order of the combined score

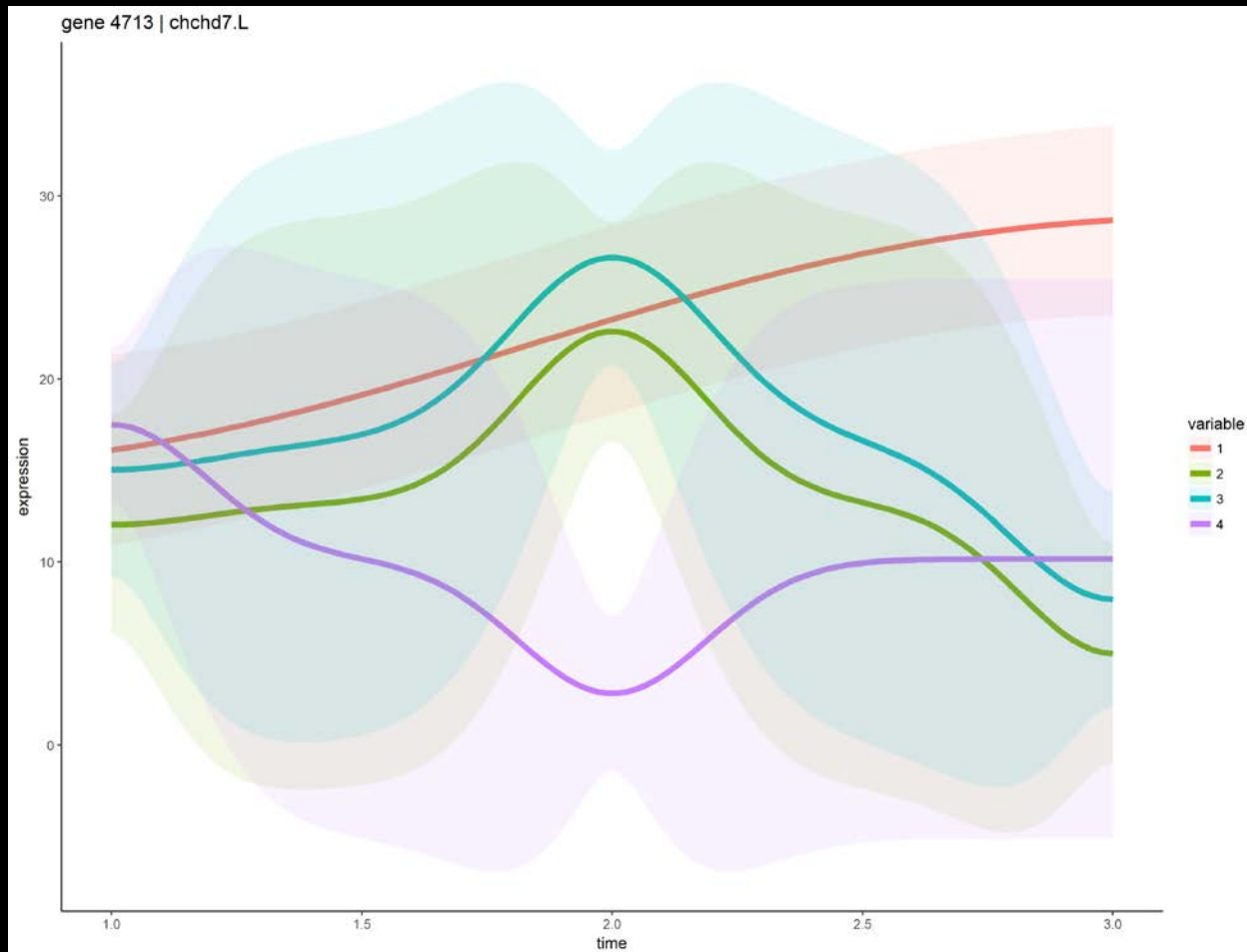- (But we present results of Euclidean too, separately...)
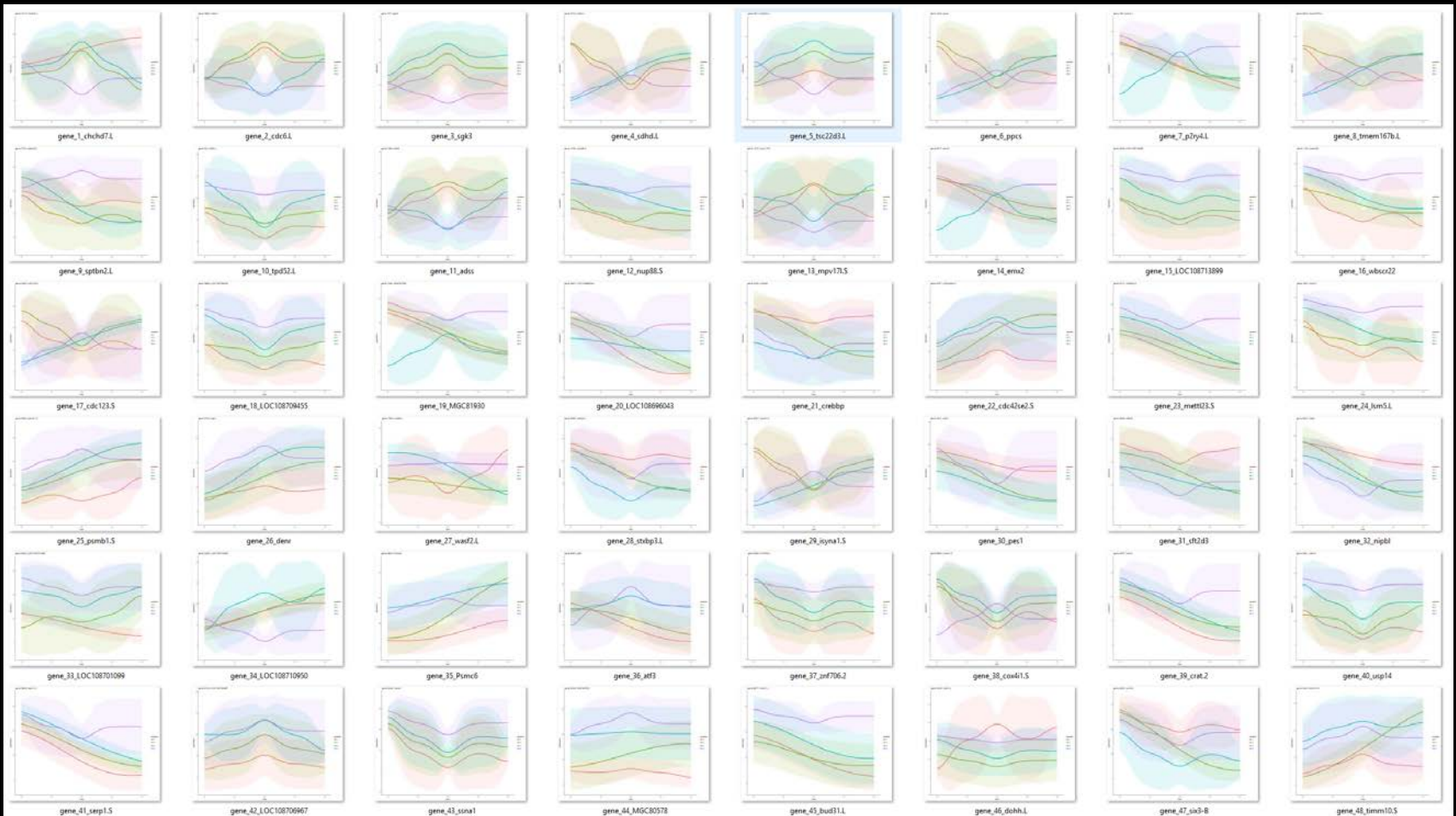
Example: Euclidean Only

Examples: Euclidean Only

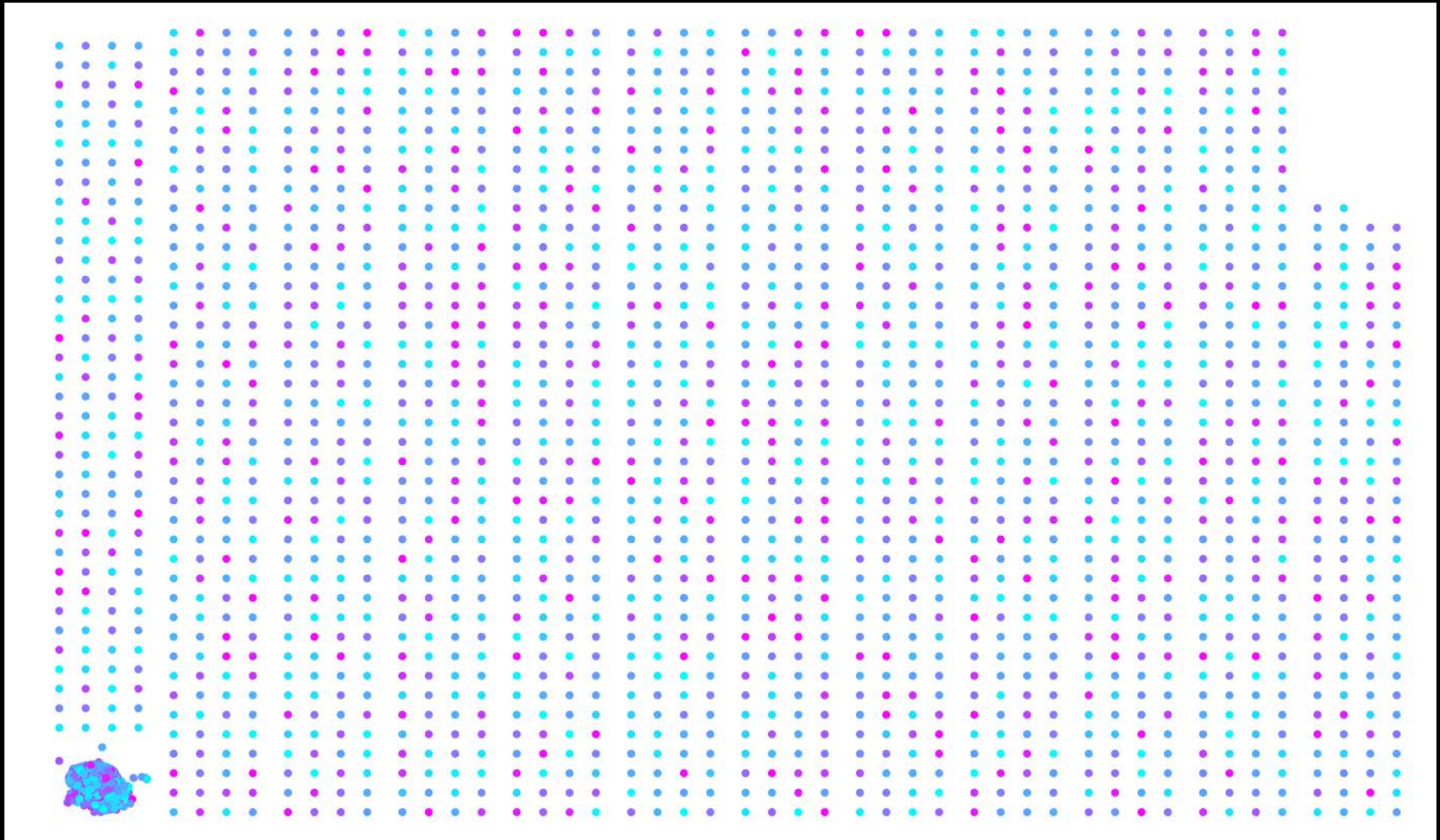# Example: Combo

Examples: Combo

Definition of Gene Groups

- We want to "group" those genes that provide a "similar separation" of the four conditions
  - Conditions have similar pairwise distances
  - Conditions are similarly informative of one another
- We use the two metrics to define a "feature space" of genes, and turn this into a clustering problem
  - "Distance over distances"
  - Can do regular hierarchical clustering on this!
  - Scale issues?
    - MI is scale-invariant
    - Distance is normalized such that maximum distance within a gene is unity
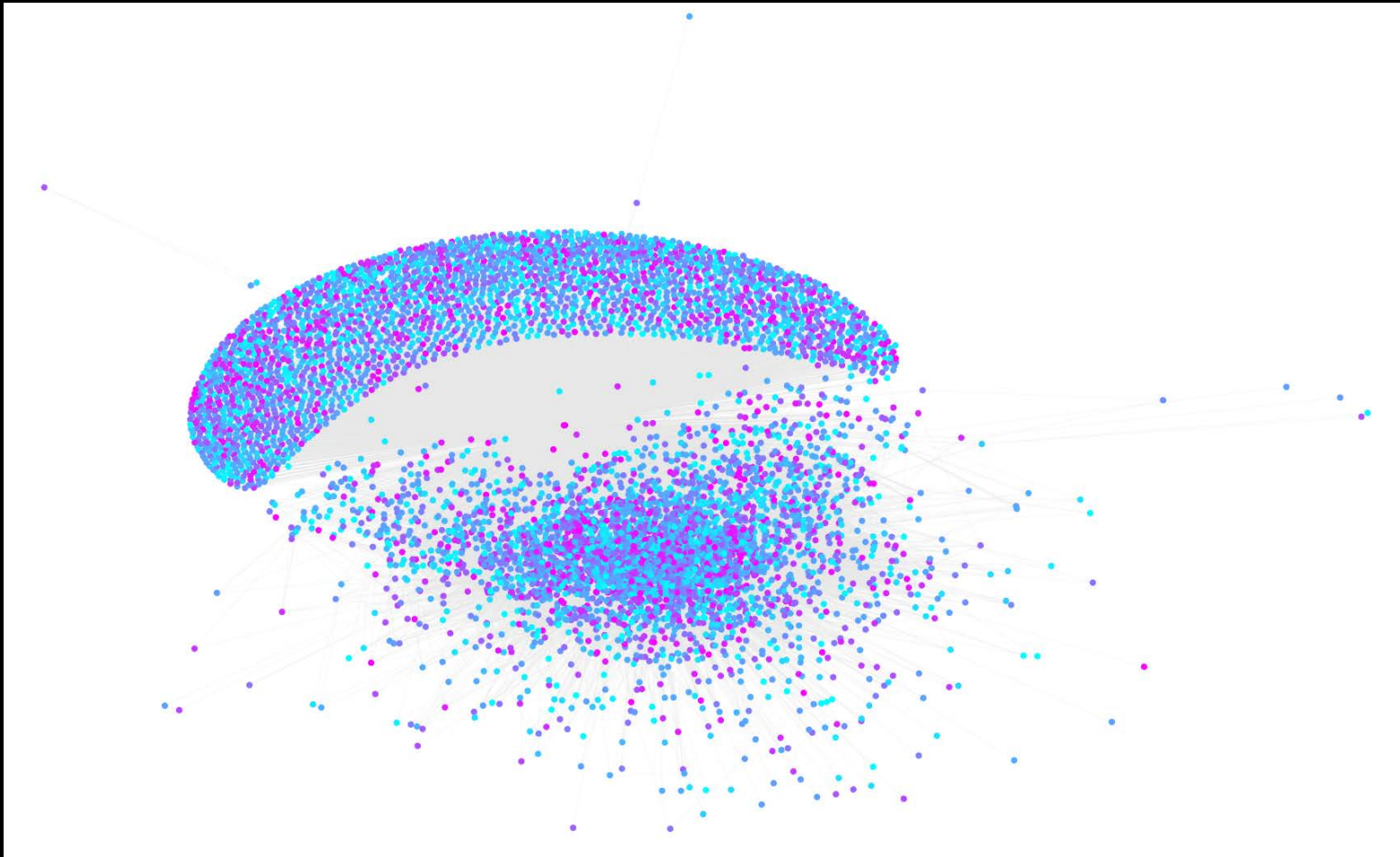
Hierarchical Clustering of Genes

- We experiment with different cluster sizes: 2, 4, 42, ...

- How do we diagnose the goodness of this clustering?

  – Use the GRN of Xenopus to define ground truth relationships between genes

  – Do we observe a topological smoothness of gene labels?

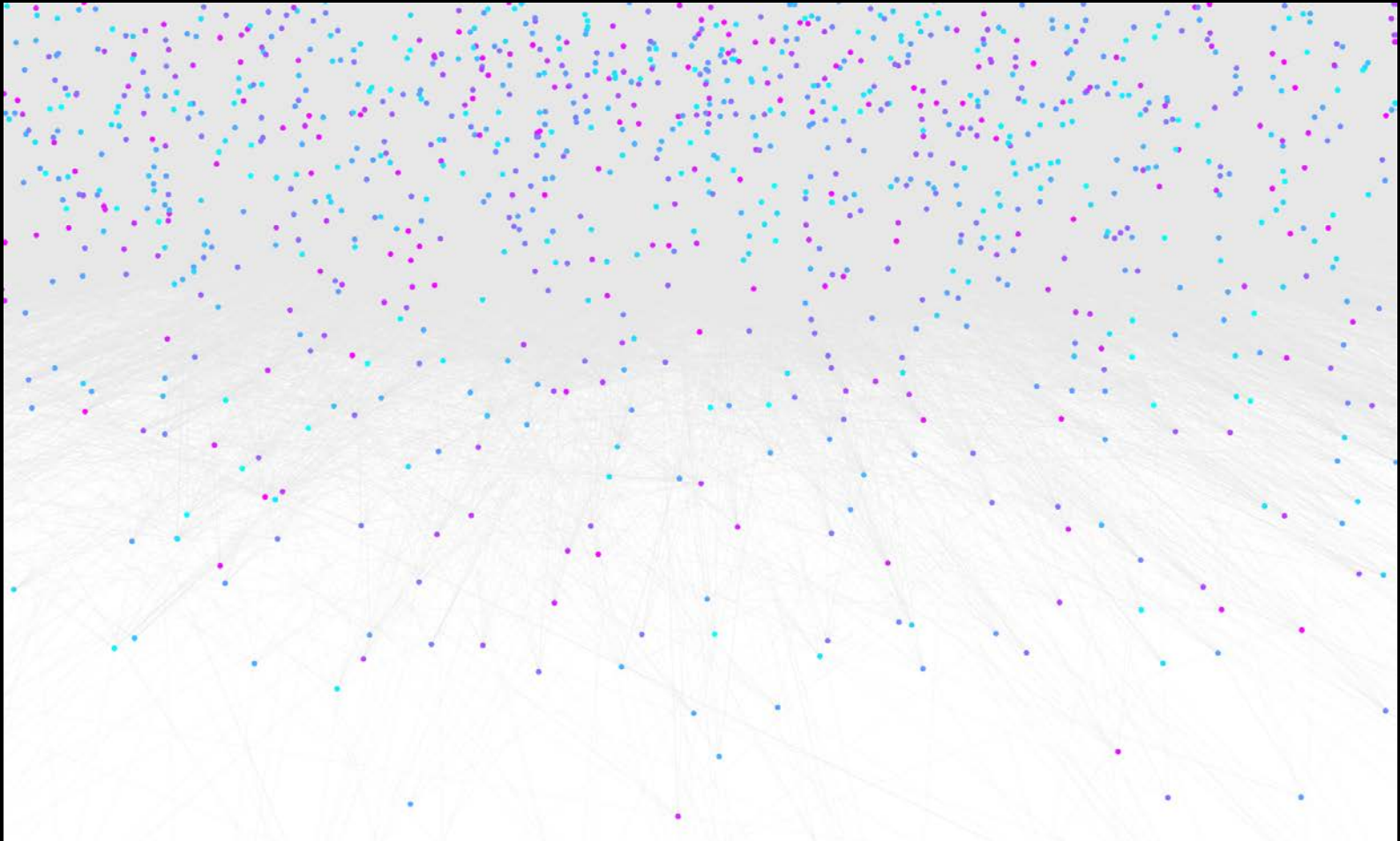   - Intuition: genes closer in regulation would be in the same group

Xenopus GRN Visualized
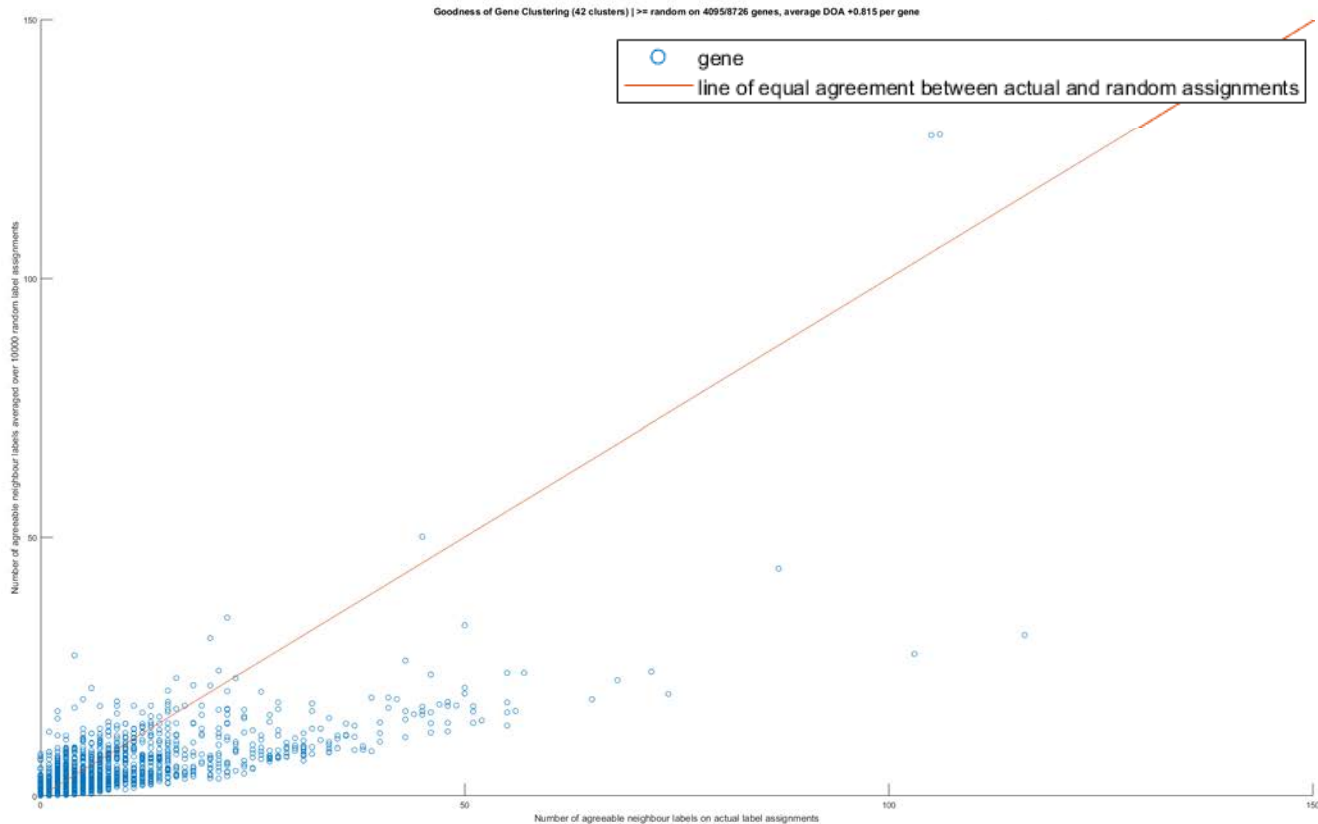
Xenopus GRN Visualized
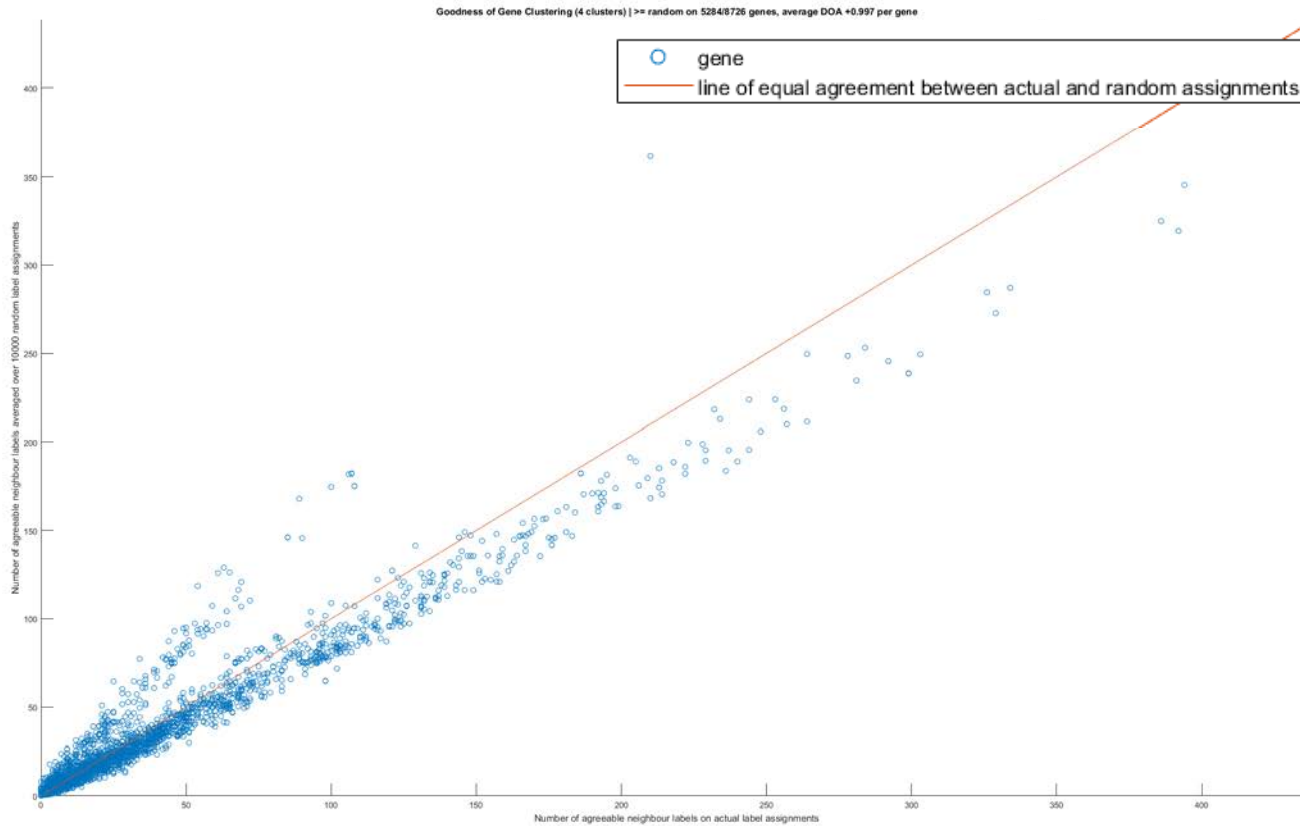
Xenopus GRN Visualized

Clustering Goodness Metric

- = Number of agreements (NOA) in the neighborhood of a gene

- We should be better than chance (random)

- We plot the NOA for every gene in
  - (1) actual, versus
  - (2) mean of 10K random assignments

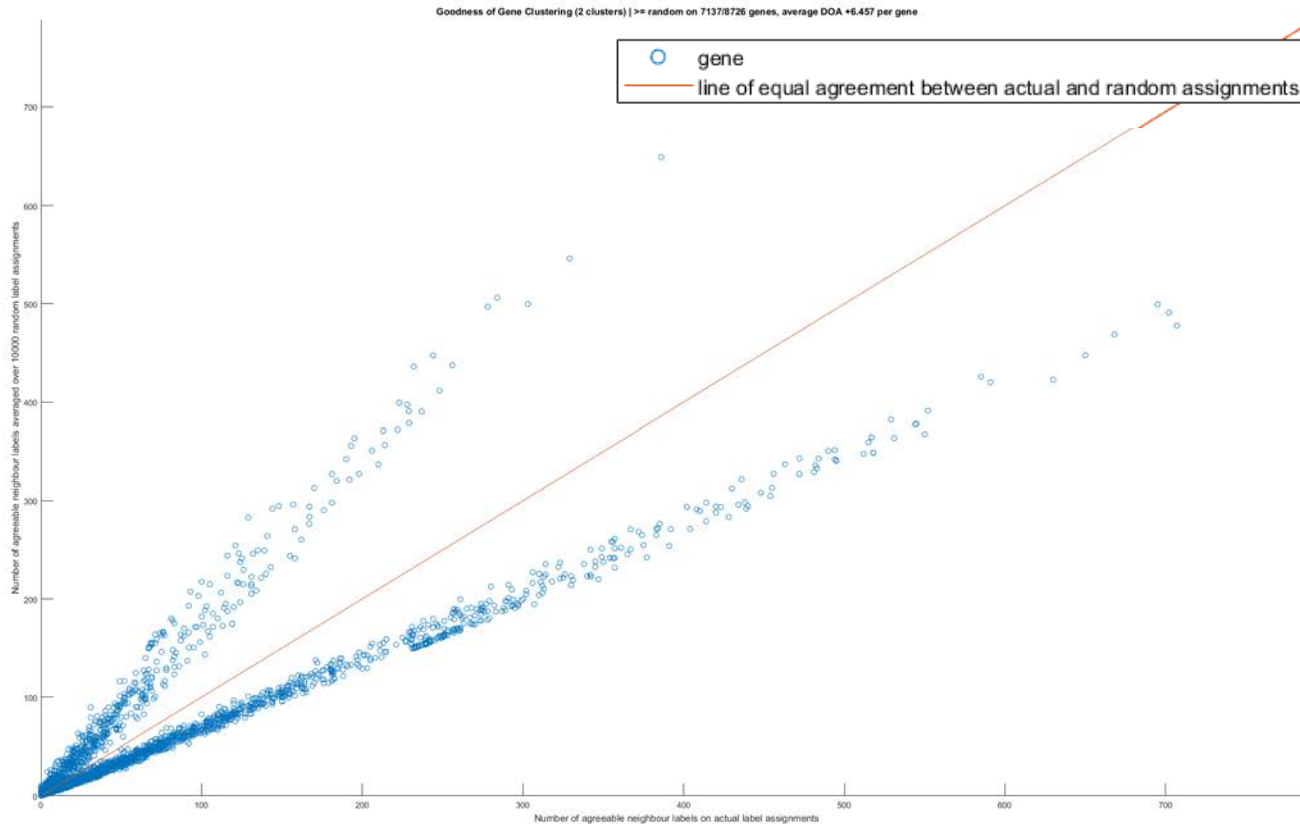- Difference of Agreements: the more positive, the better
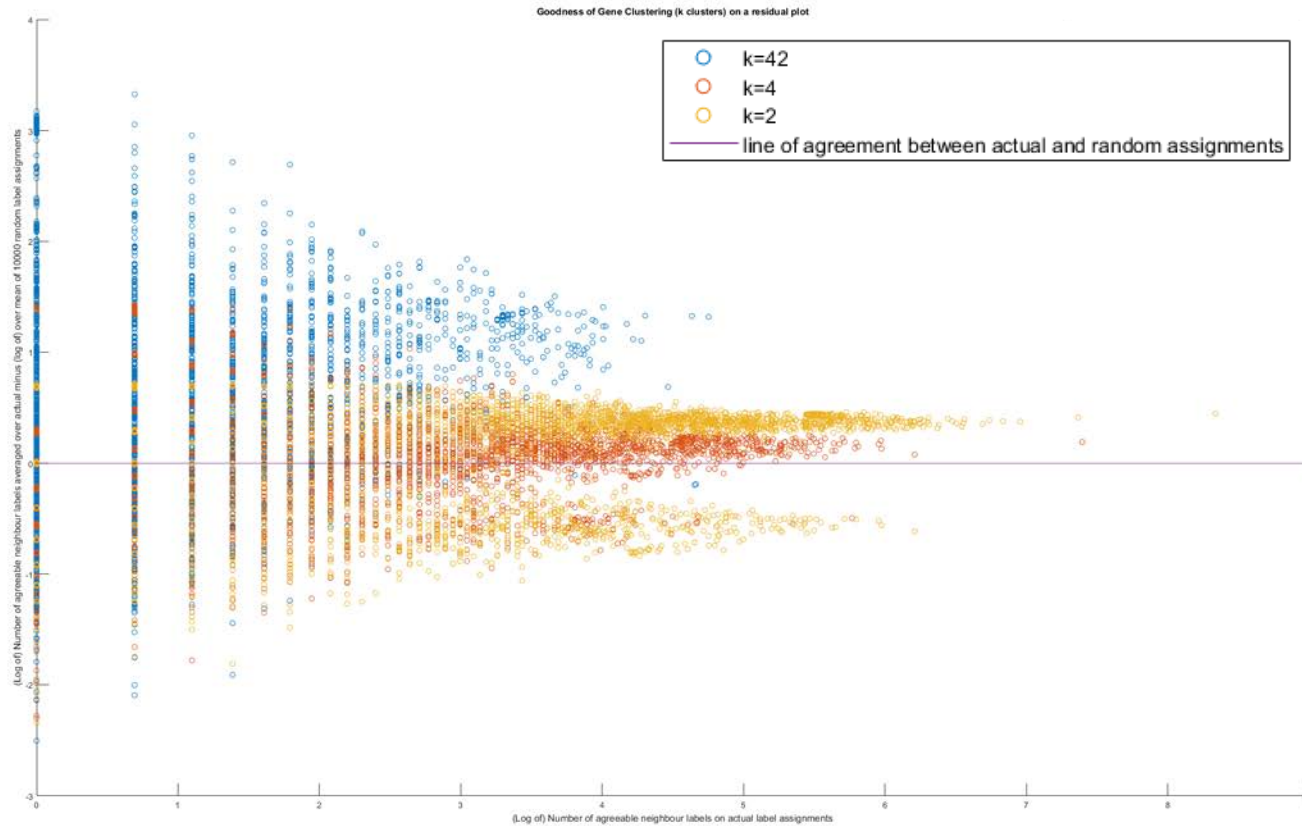
Clustering Goodness k=42 (DOA +0.815 per gene)

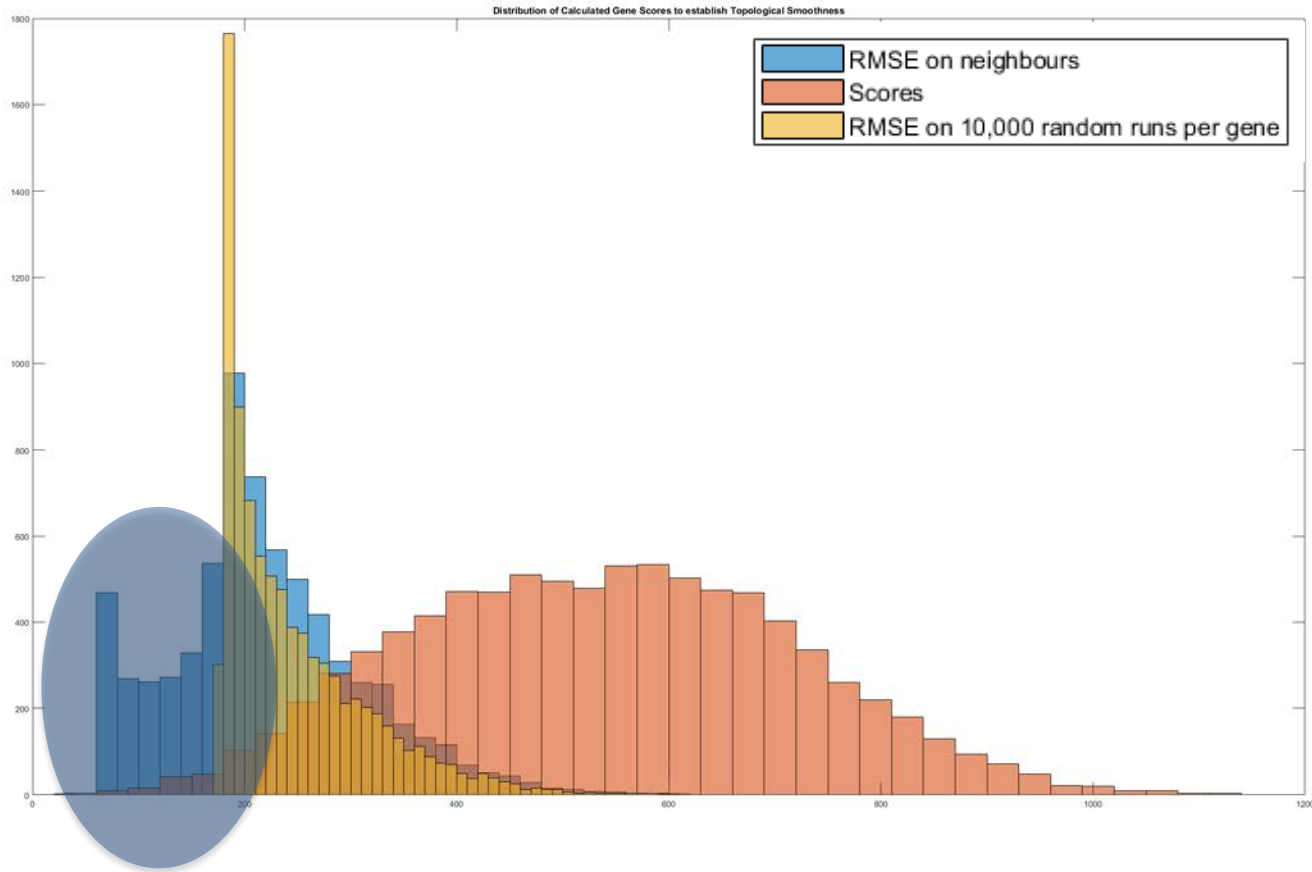Goodness of Gene Clustering (42 clusters) | >= random on 4095/8726 genes, average DOA +0.815 per gene

Clustering Goodness k=4 (DOA +0.997 per gene)

Clustering Goodness k=2
(DOA +6.457 per gene)

Goodness of Gene Clustering (2 clusters) | >= random on 7137/8726 genes, average DOA +6.457 per gene

○ gene
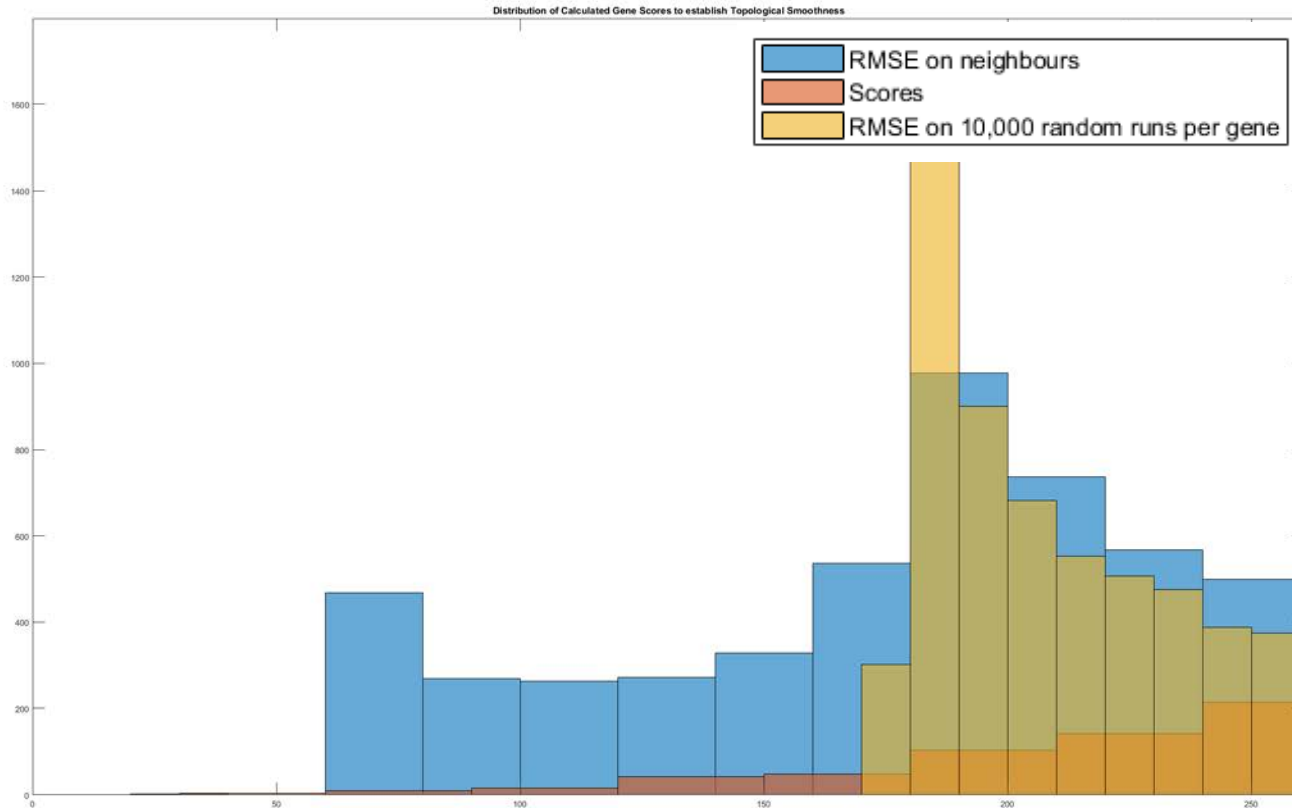— line of equal agreement between actual and random assignments

Clustering Goodness (Log-Log plot of DOA)

Are the Gene Scores also topologically smooth?

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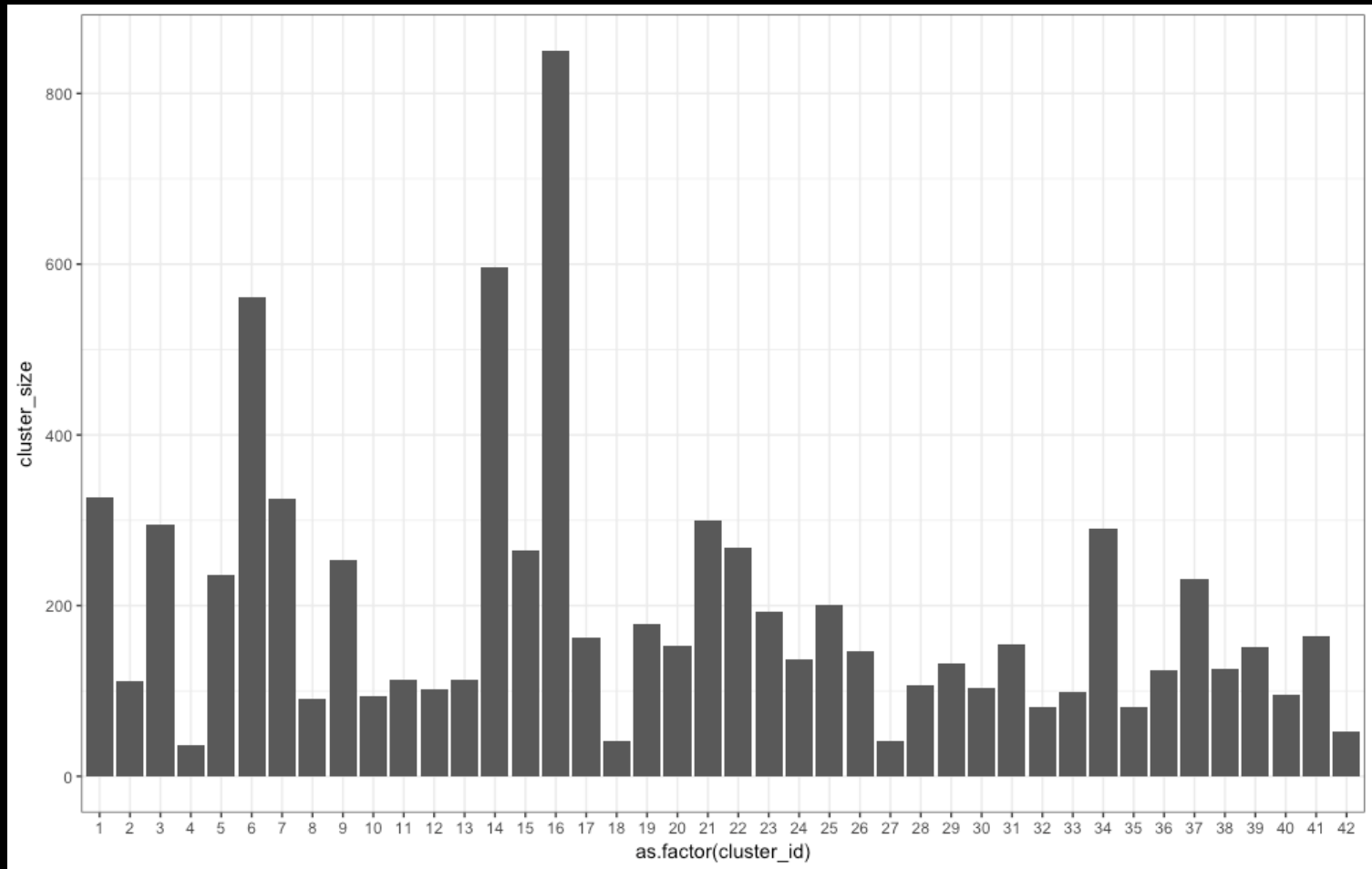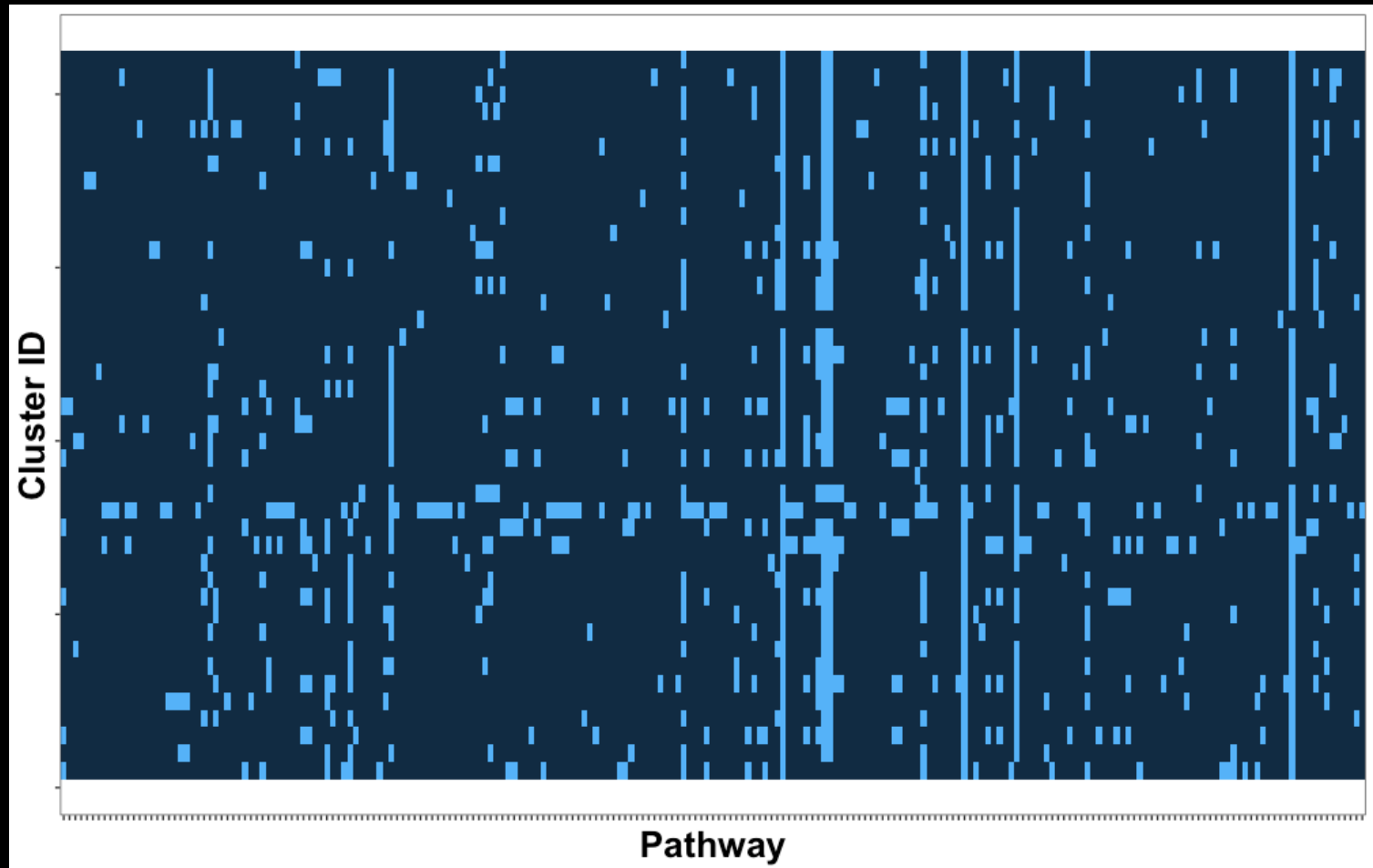# Are the Gene Scores also topologically smooth?
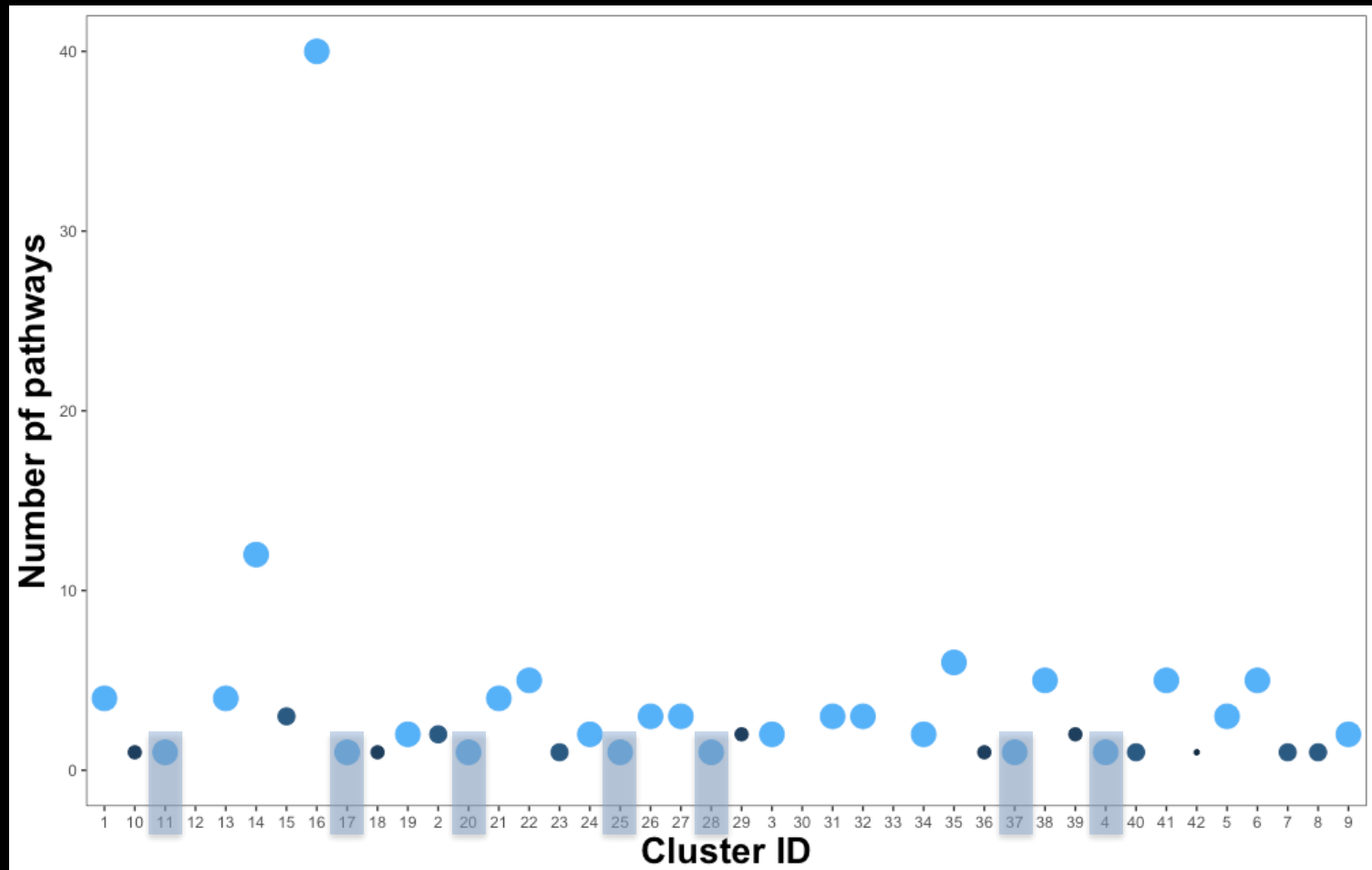## (GRN has 504268 true edges out of 38067175; AUROC=0.501)

ROC curve assuming Gene Distances to define a Gene Regulatory netowrk for frogs (8726 genes, 504268/38067175 true edges), AUROC=0.501

Mapping Gene Groups to Pathways

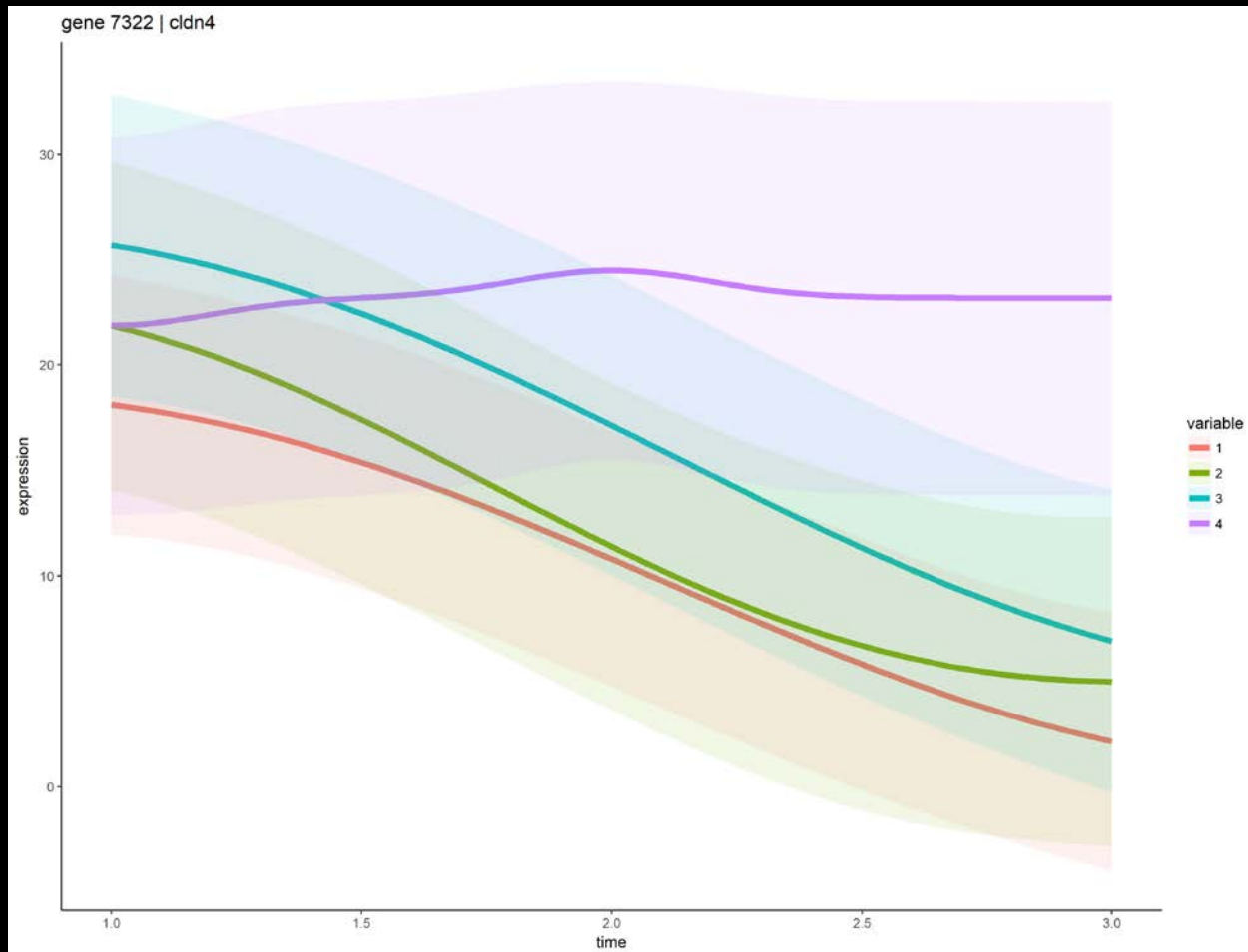# Mapping Gene Groups to Pathways

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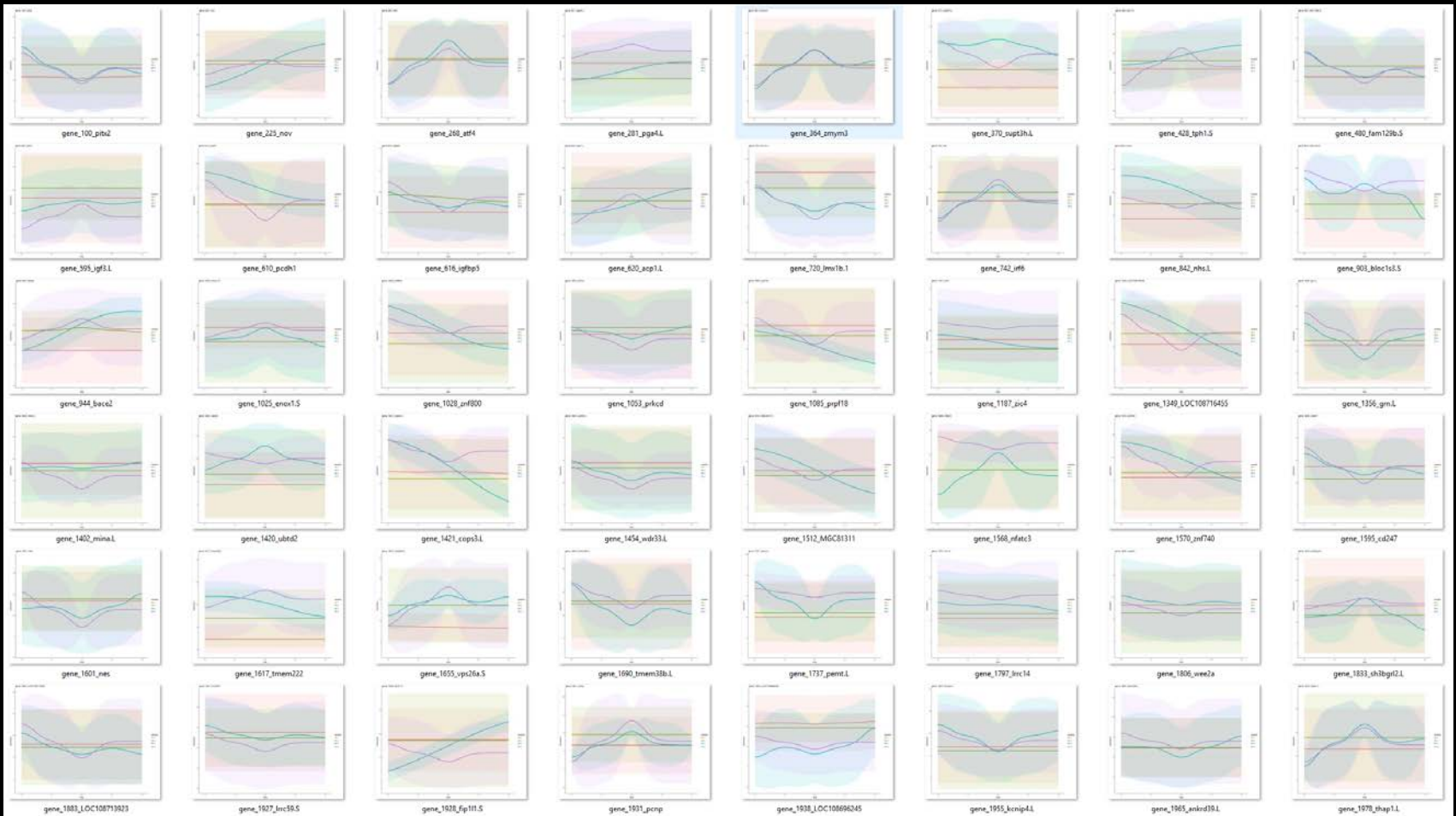| Gene Group	Mapped Pathway
4	g2/m_dna_replication_checkpoint
11	rna_polymerase_ii_transcription
17	dcc_mediated_attractive_signaling
20	**abacavir_transport_and_metabolism**
25	norc_negatively_regulates_rrna_expression
28	glycogen_synthesis
37	**scavenging_of_heme_from_plasma**

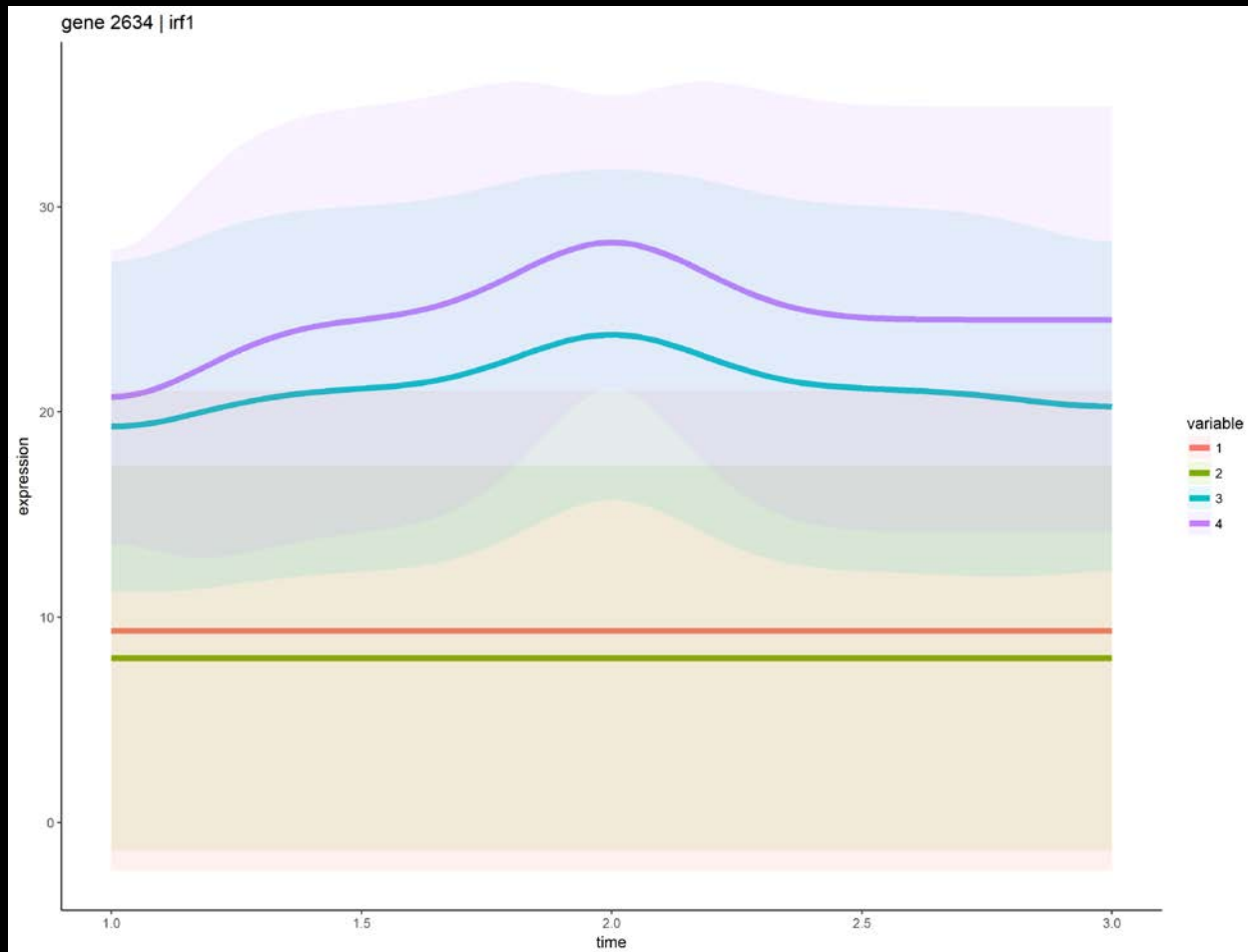# Gene Group 20 (abacavir)

Example Gene from Group 20

gene 7322 | cldn4

Gene Group 37 (heme)

Example Gene from Group 37
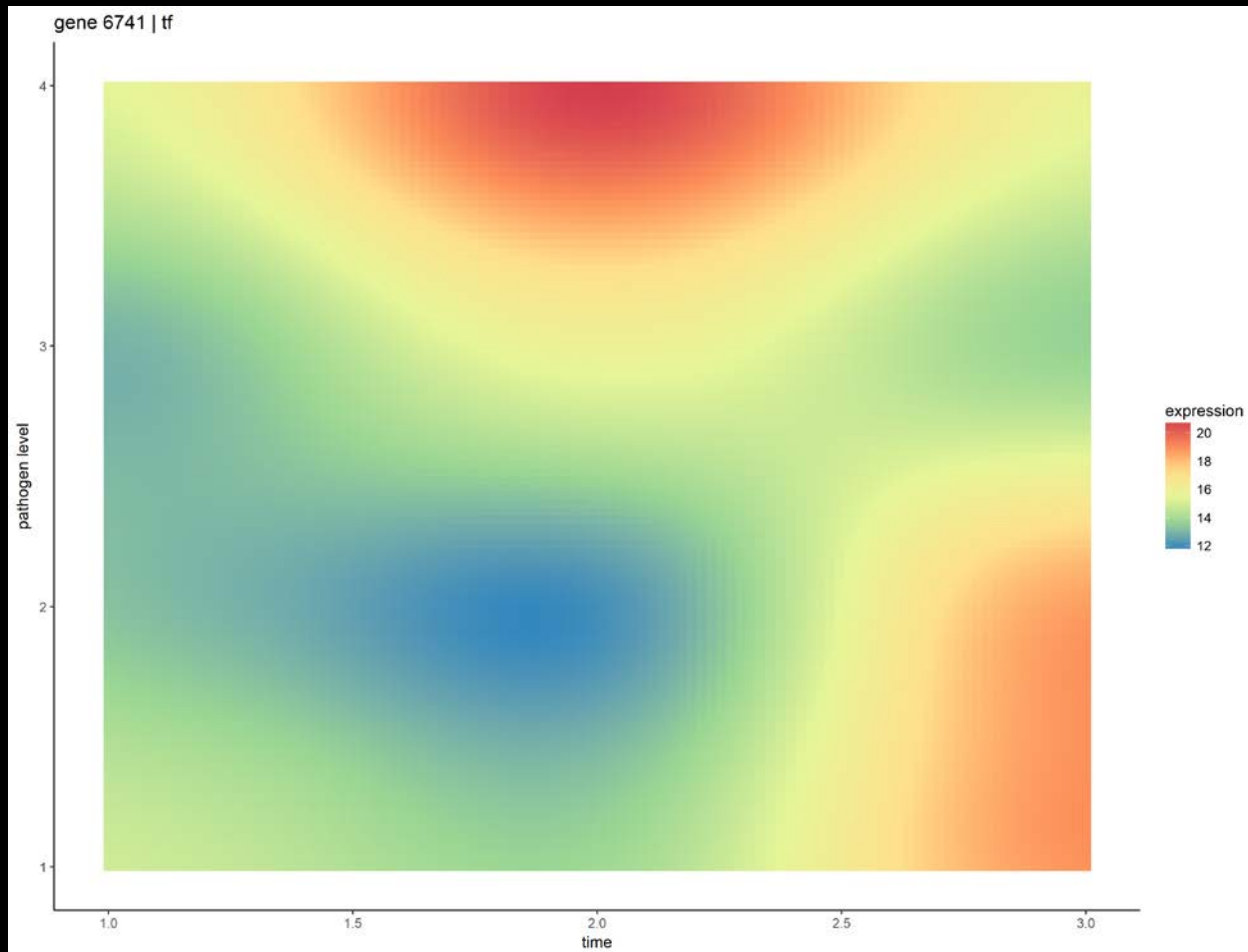
GPR for Omics, Method 2

- Treat every gene as an independent GP mapping from   time   space to expression space
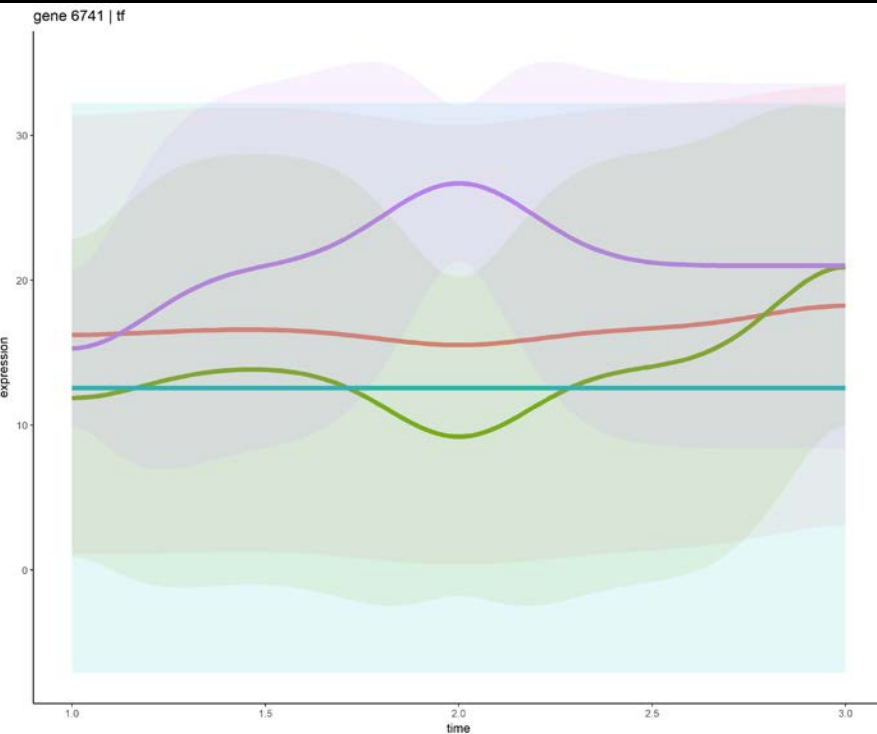
$$f: t \quad \rightarrow g$$
$$f(t' \quad) \sim GP\big(m(t \quad), k(t, \quad t' \quad)\big) + \beta$$

- Every gene has only 1 GP model to be learnt

- Allows regression over both time and pathogen levels, together
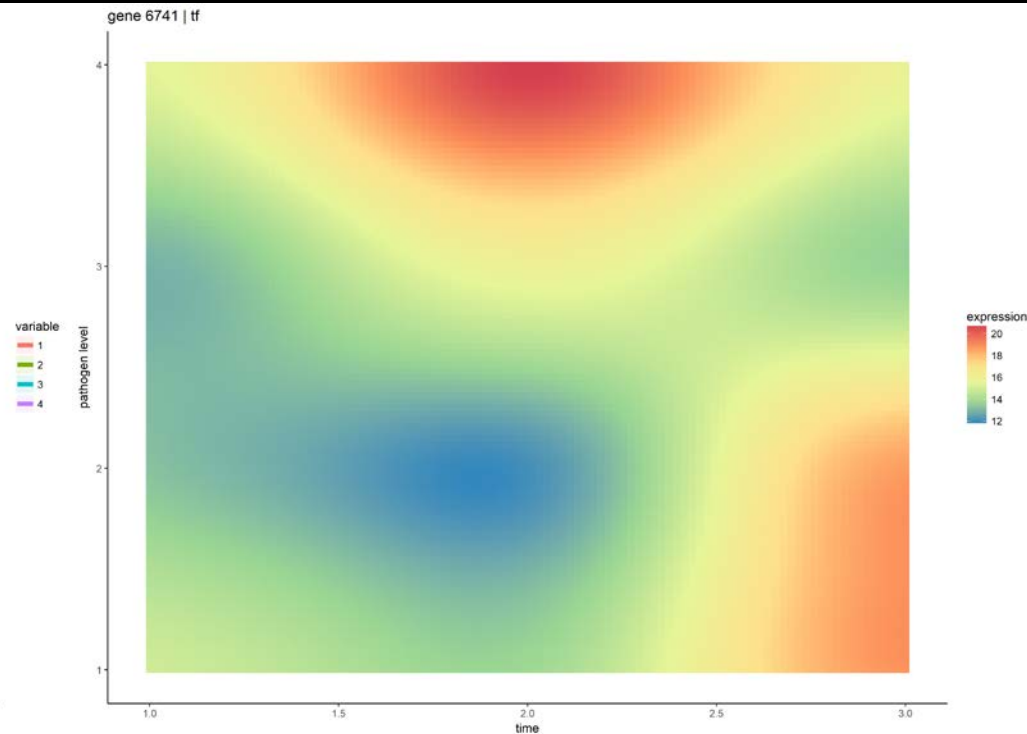
Example: Transferrin (probe1)
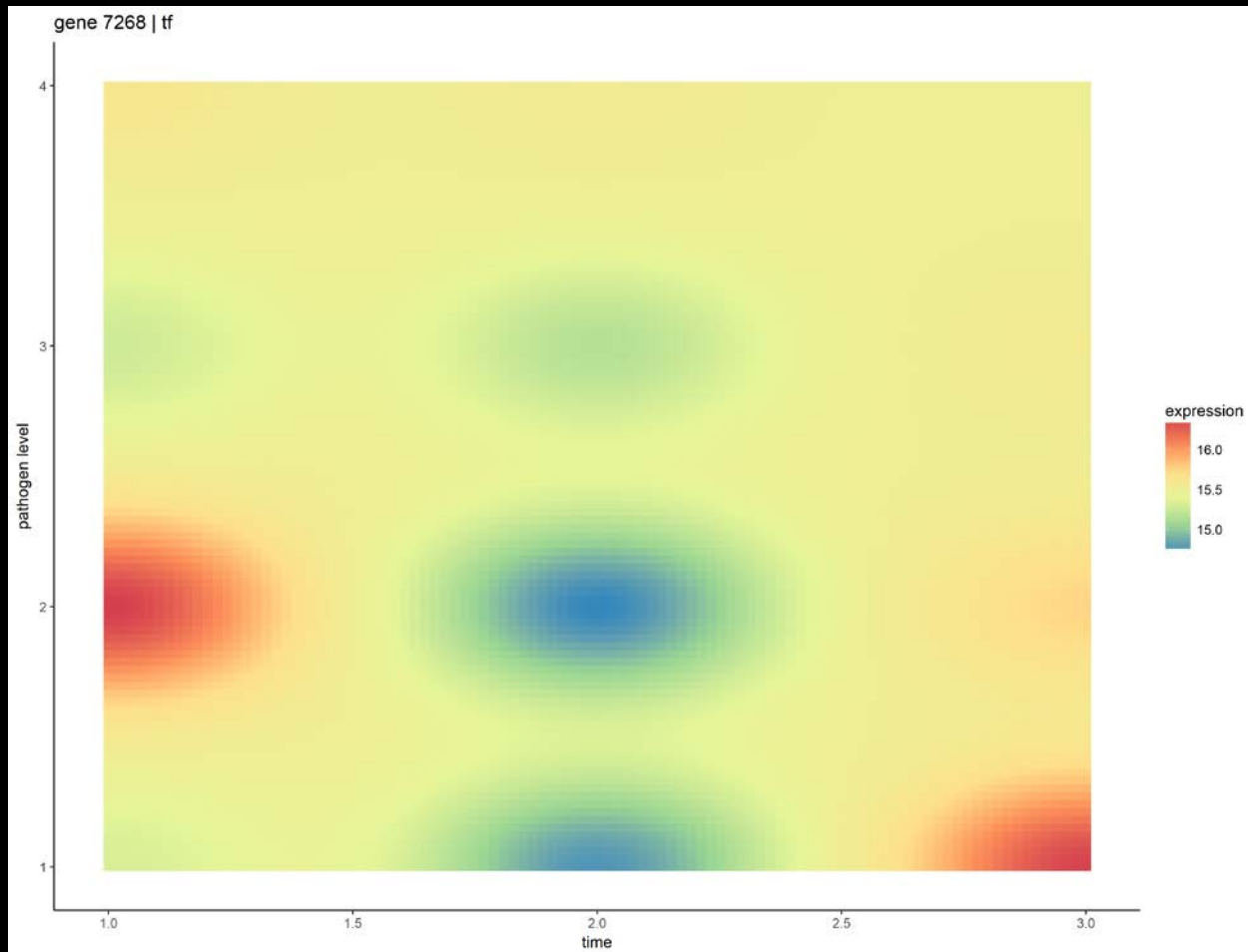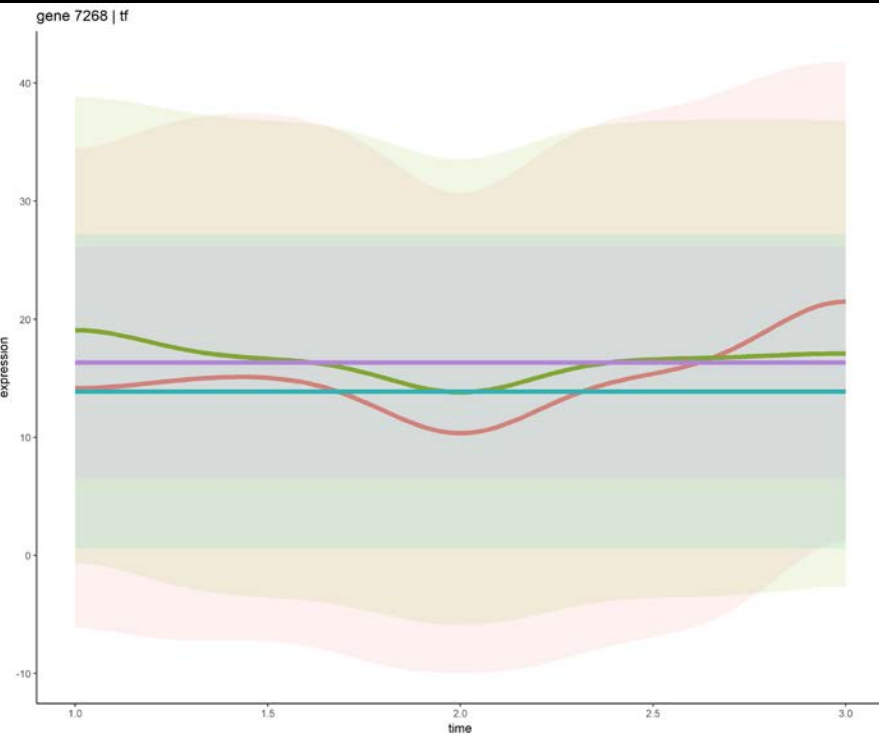
Example: Transferrin (probe1)

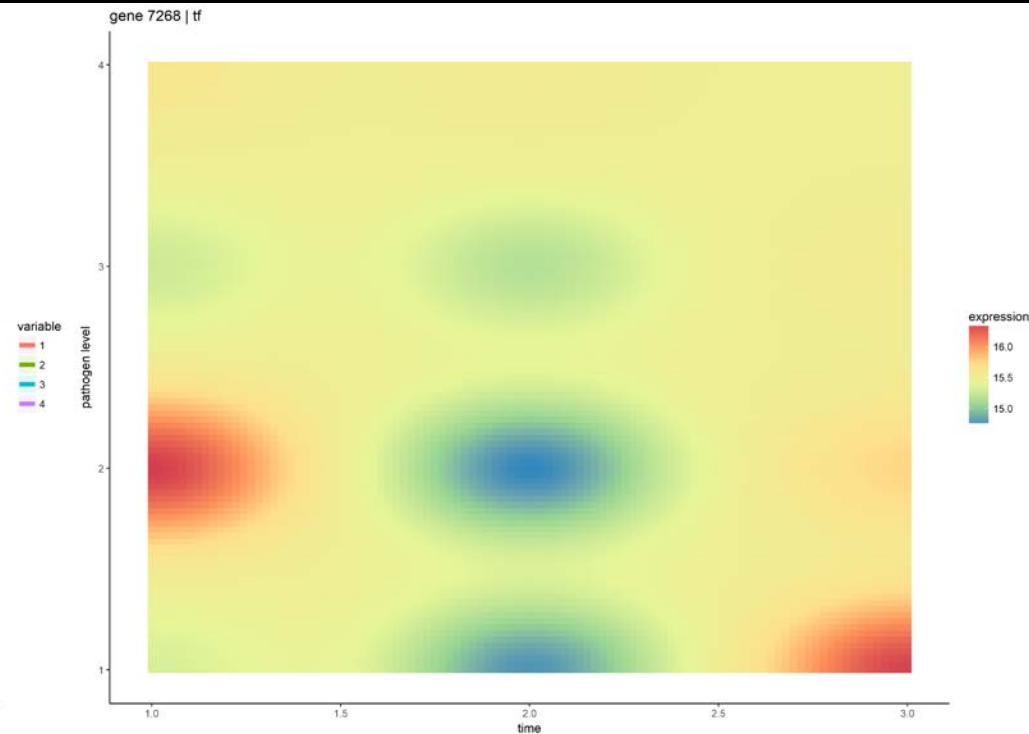Univariate method

Bivariate method

Example: Transferrin (probe2)

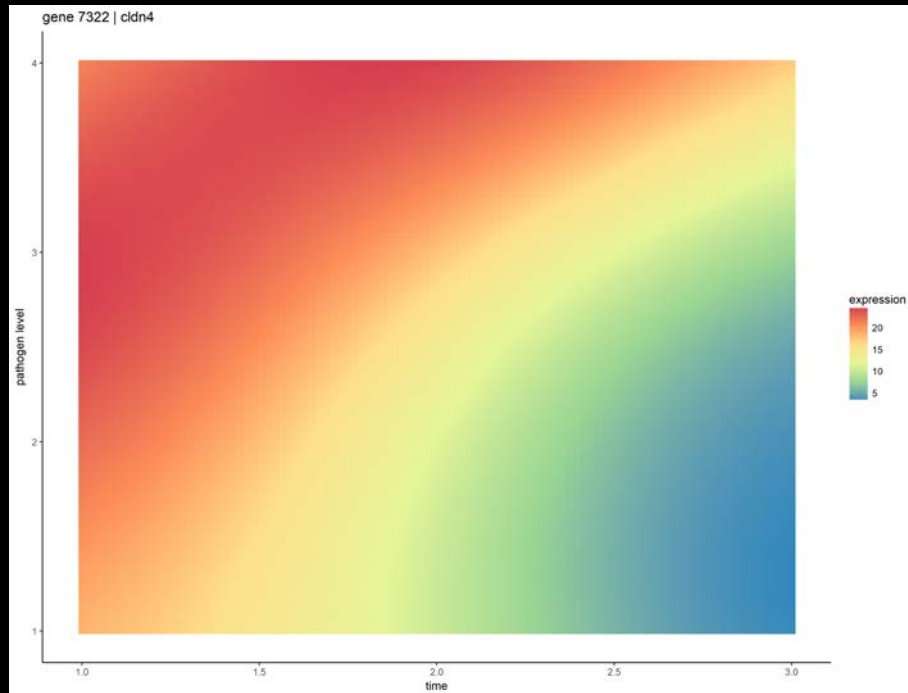# Example: Transferrin (probe2)
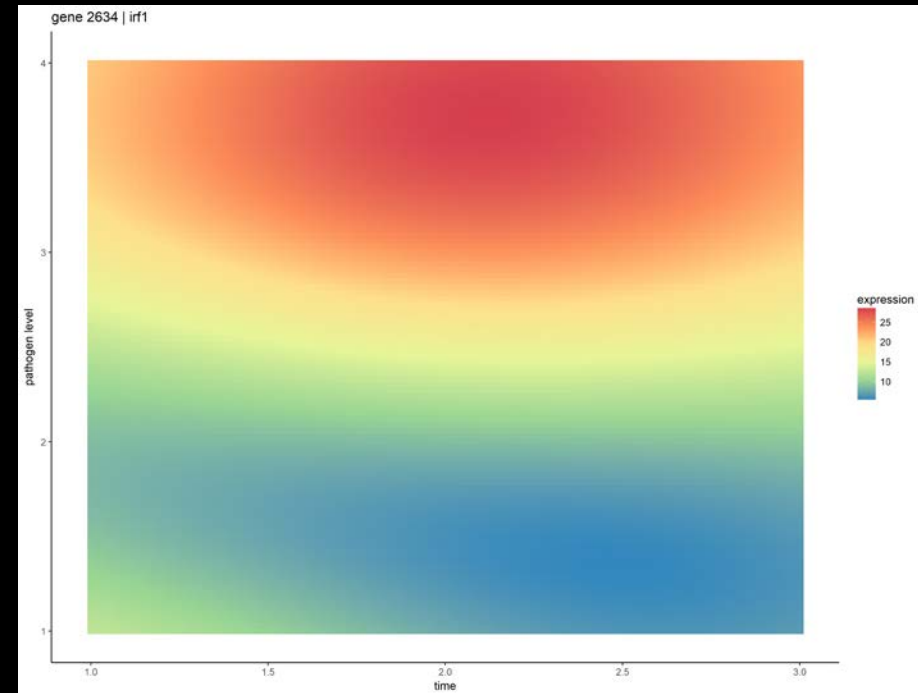

Univariate method

Bivariate method

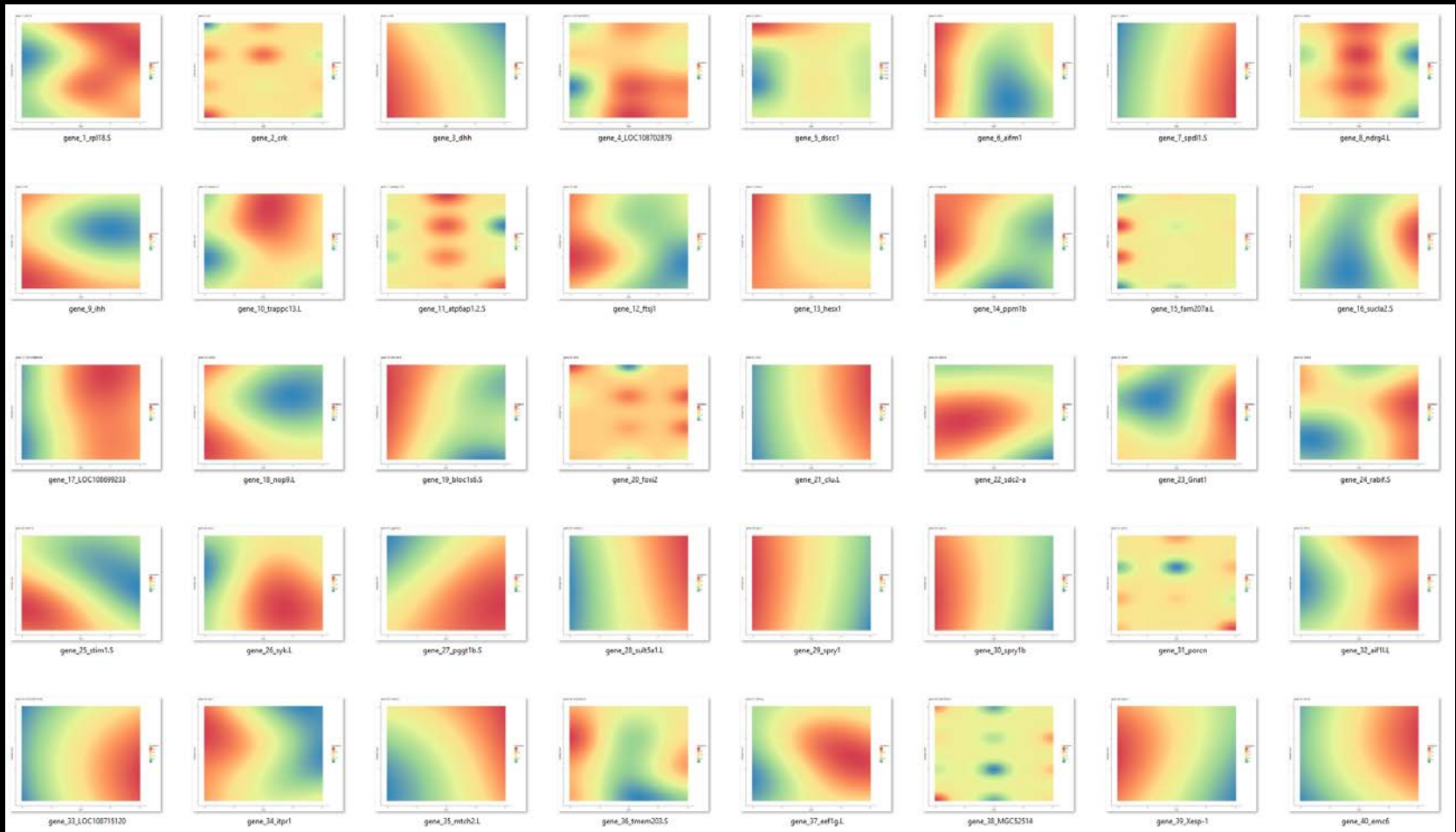# Examples: from abacavir and heme Gene Groups
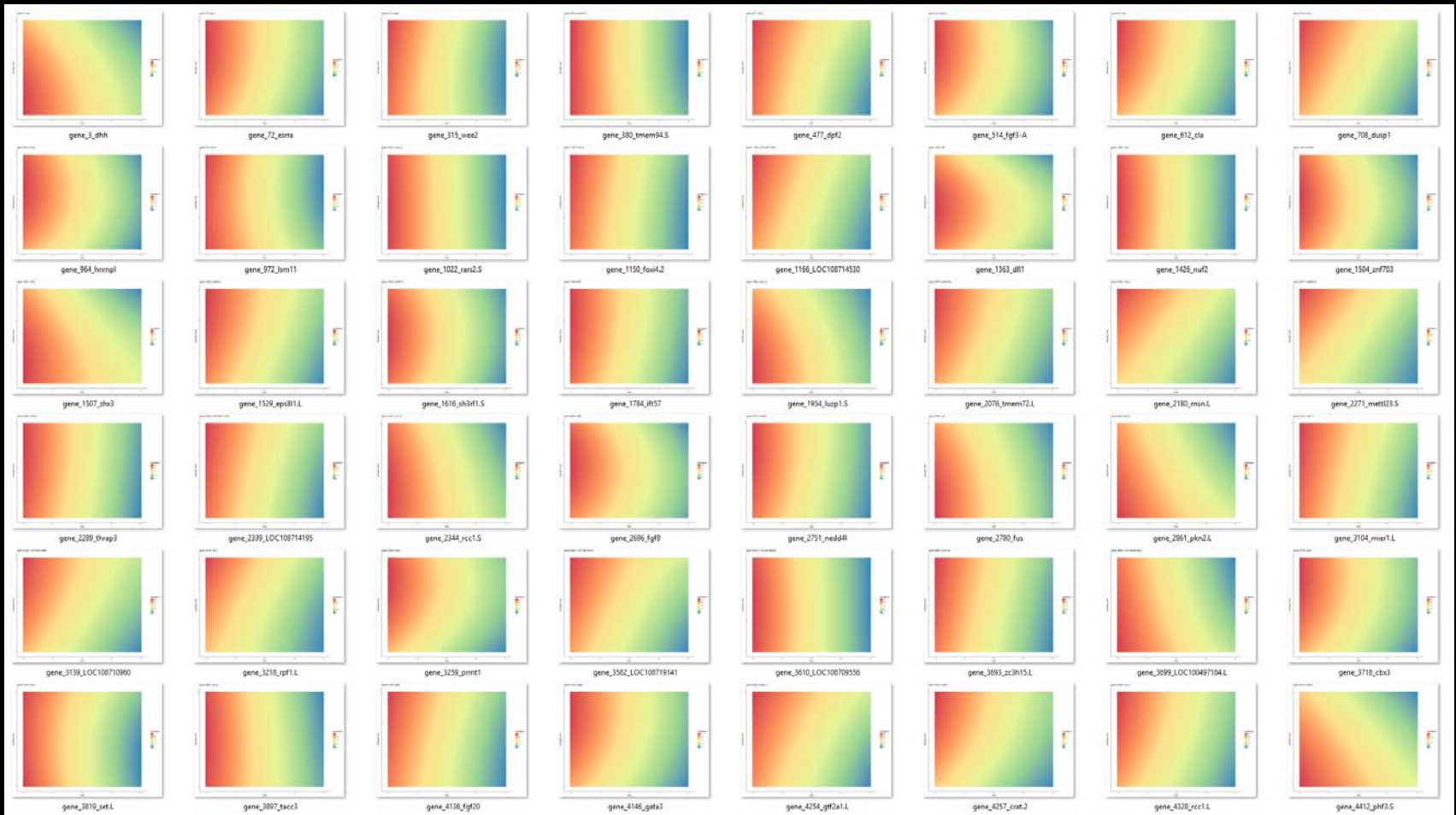

abacavir gene group

heme gene group

For all 8726 genes…

To extract key genes and gene groups...
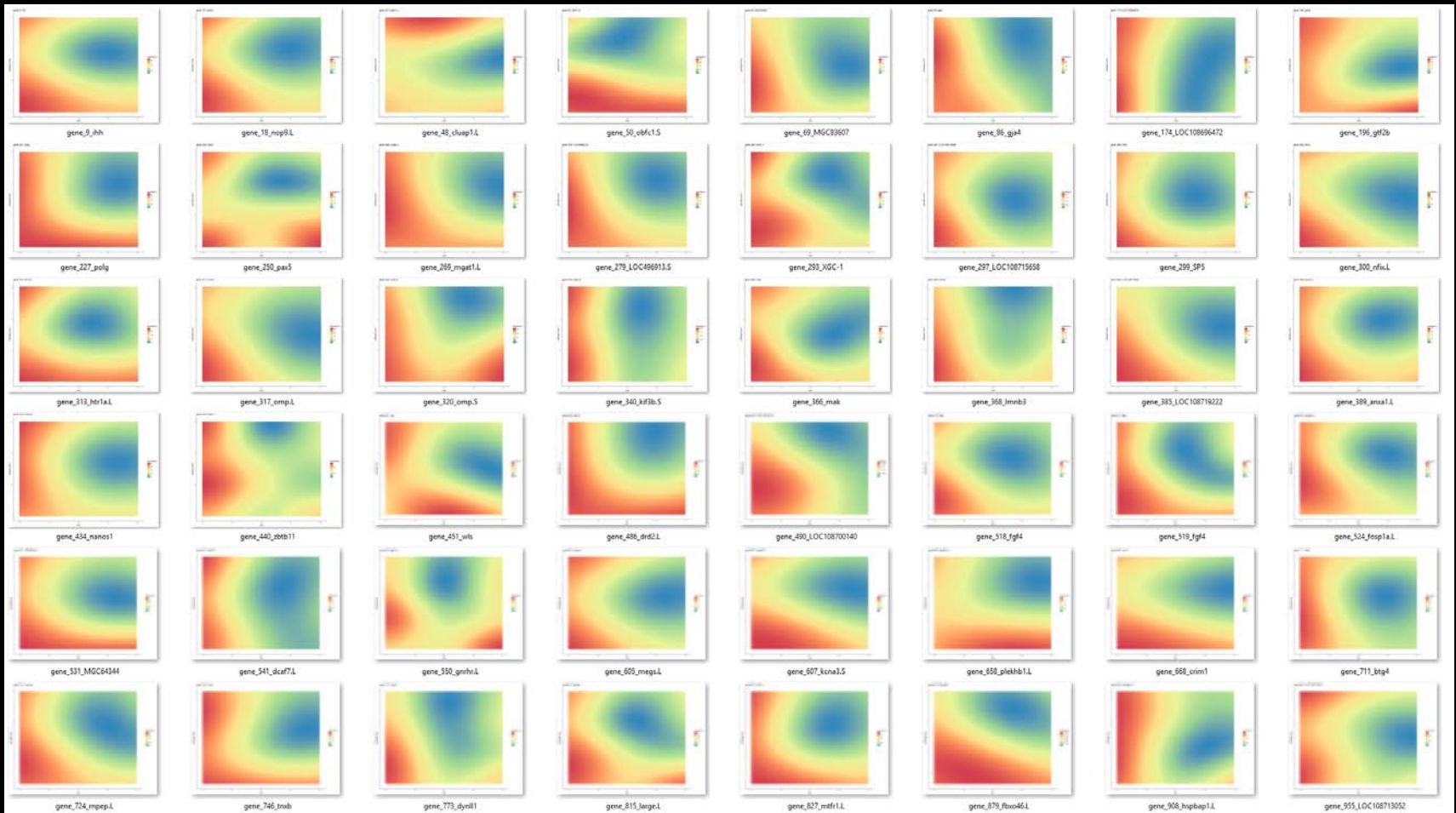
- Visually!

- Question: How do we compare two GP models?
  - Can compare in model space itself because we have same inputs to all GP models!

- Solution: we use the mean function and covariance function of learnt GP model to define a "feature space" of genes
  - We conveniently manage to ignore scale and bias in comparing genes

Examples: Gene Group

Examples: Gene Group

Summary

- Model(s) for time-series data under multiple conditions, with limited number of samples

Data Points → (Conditional Univariate) GP Models → Smooth Mean Time Trajectories → Gene Groups

Data Points → (Joint Bivariate) GP Model → Gene Groups
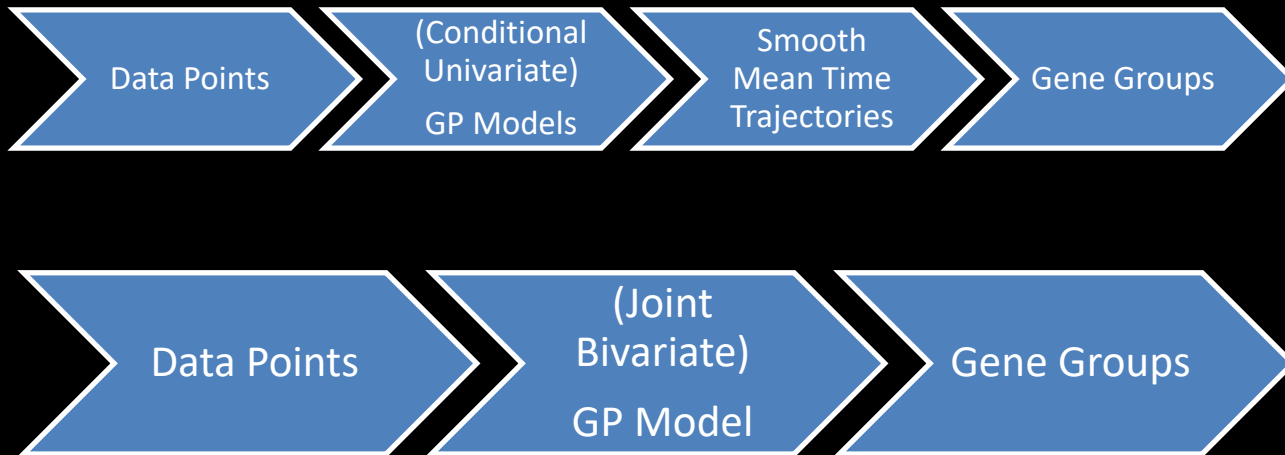
Summary

- Model(s) for time-series data under multiple conditions, with limited number of samples

- Good tool for quick visual inspections

- Outputs key genes and gene groups to look at, mapped to appropriate pathways

- Generalized to any temporal dataset with multiple conditions where "smooth" assumption can be applied

- Next steps:
  - Use actual CFU counts of every sample in the joint bivariate regression model
  - Build a simple browser-based app for everyone to use